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August 04, 2005

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APPLICATION NUMBER: 60/625,871

FILING DATE: *November 08, 2004*

RELATED PCT APPLICATION NUMBER: *PCT/US05/11626*



Certified by

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PROVISIONAL APPLICATION COVER SHEET
Additional Page

PTO/SB/16 (09-04)

Approved for use through 07/31/2006. OMB 0651-0032

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| First Named Inventor | Joseph R. Garlich | Docket Number 01656.0011.PZUS00 |
|--|-------------------|---|
| INVENTOR(S)/APPLICANT(S) | | |
| Given Name (first and middle [if any]) | Family or Surname | Residence (City and either State or Foreign Country) |
| Xiaodong | Peng | |
| Jin | Su | |
| Tim C. | Smith | |
| | | |

Number 1 of 2

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PTO/SB/17 (10-04v2)
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FEE TRANSMITTAL for FY 2005

Effective 10/01/2004. Patent fees are subject to annual revision.

☒ Applicant claims small entity status. See 37 CFR 1.27

TOTAL AMOUNT OF PAYMENT (\$) 80.00

Complete if Known

| | |
|----------------------|-------------------|
| Application Number | Not yet assigned |
| Filing Date | November 8, 2004 |
| First Named Inventor | Joseph R. Garlich |
| Examiner Name | Not yet assigned |
| Art Unit | Not yet assigned |
| Attorney Docket No. | 01656.0011.PZUS00 |

METHOD OF PAYMENT (check all that apply)

☐ Check ☐ Credit card ☐ Money Order ☐ Other ☐ None

☒ Deposit Account:

Deposit Account Number: 08-3038
Deposit Account Name: Howrey Simon Arnold & White LLP

The Director is authorized to: (check all that apply)

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FEE CALCULATION

1. BASIC FILING FEE

| Large Entity Fee Code (\$) | Small Entity Fee Code (\$) | Fee Description | Fee Paid |
|----------------------------|----------------------------|------------------------|-------------|
| 1001 790 | 2001 395 | Utility filing fee | |
| 1002 350 | 2002 175 | Design filing fee | |
| 1003 550 | 2003 275 | Plant filing fee | |
| 1004 790 | 2004 395 | Reissue filing fee | |
| 1005 160 | 2005 80 | Provisional filing fee | 80.00 |
| SUBTOTAL (1) | | | (\$) 80.00 |

2. EXTRA CLAIM FEES FOR UTILITY AND REISSUE

| Total Claims | Extra Claims | Fee from below | Fee Paid |
|--------------------|--------------|----------------|----------|
| Independent Claims | -20** = | X | |
| Multiple Dependent | -3** = | X | |

| Large Entity Fee Code (\$) | Small Entity Fee Code (\$) | Fee Description |
|----------------------------|----------------------------|--|
| 1202 18 | 2202 9 | Claims in excess of 20 |
| 1201 88 | 2201 44 | Independent claims in excess of 3 |
| 1203 300 | 2203 150 | Multiple dependent claim, if not paid |
| 1204 88 | 2204 44 | ** Reissue independent claims over original patent |
| 1205 18 | 2205 9 | ** Reissue claims in excess of 20 and over original patent |

SUBTOTAL (2) (\$) -0-

**or number previously paid, if greater; For Reissues, see above

FEE CALCULATION (continued)

3. ADDITIONAL FEES

| Large Entity Fee Code (\$) | Small Entity Fee Code (\$) | Fee Description | Fee Paid |
|----------------------------|----------------------------|--|----------|
| 1051 130 | 2051 65 | Surcharge - late filing fee or oath | |
| 1052 50 | 2052 25 | Surcharge - late provisional filing fee or cover sheet | |
| 1053 130 | 1053 130 | Non-English specification | |
| 1812 2,520 | 1812 2,520 | For filing a request for <i>ex parte</i> reexamination | |
| 1804 920* | 1804 920* | Requesting publication of SIR prior to Examiner action | |
| 1805 1,840* | 1805 1,840* | Requesting publication of SIR after Examiner action | |
| 1251 110 | 2251 55 | Extension for reply within first month | |
| 1252 430 | 2252 215 | Extension for reply within second month | |
| 1253 980 | 2253 490 | Extension for reply within third month | |
| 1254 1,530 | 2254 765 | Extension for reply within fourth month | |
| 1255 2,080 | 2255 1,040 | Extension for reply within fifth month | |
| 1401 340 | 2401 170 | Notice of Appeal | |
| 1402 340 | 2402 170 | Filing a brief in support of an appeal | |
| 1403 300 | 2403 150 | Request for oral hearing | |
| 1451 1,510 | 1451 1,510 | Petition to institute a public use proceeding | |
| 1452 110 | 2452 55 | Petition to revive - unavoidable | |
| 1453 1,370 | 2453 685 | Petition to revive - unintentional | |
| 1501 1,370 | 2501 685 | Utility issue fee (or reissue) | |
| 1502 490 | 2502 245 | Design issue fee | |
| 1503 660 | 2503 330 | Plant issue fee | |
| 1460 130 | 1460 130 | Petitions to the Commissioner | |
| 1807 50 | 1807 50 | Processing fee under 37 CFR 1.17(q) | |
| 1806 180 | 1806 180 | Submission of Information Disclosure Stmt | |
| 8021 40 | 8021 40 | Recording each patent assignment per property (times number of properties) | |
| 1809 790 | 2809 395 | Filing a submission after final rejection (37 CFR 1.129(a)) | |
| 1810 790 | 2810 395 | For each additional invention to be examined (37 CFR 1.129(b)) | |
| 1801 790 | 2801 395 | Request for Continued Examination (RCE) | |
| 1802 900 | 1802 900 | Request for expedited examination of a design application | |

Other fee (specify)

*Reduced by Basic Filing Fee Paid

SUBTOTAL (3) (\$) -0-

SUBMITTED BY

| | | | | | |
|-------------------|----------------------------|-----------------------------------|------------------|-----------|--------------|
| Name (Print/Type) | Teddy C. Scott, Jr., Ph.D. | Registration No. (Attorney/Agent) | 53,573 | Telephone | 312 846-5621 |
| Signature | | Date | November 8, 2004 | | |

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**TRANSMITTAL
FORM**

(to be used for all correspondence after initial filing)

Total Number of Pages in This Submission

60

Application Number

Not yet assigned

Filing Date

November 8, 2004

First Named Inventor

Joseph R. Garlich

Art Unit

Not yet assigned

Examiner Name

Not yet assigned

Attorney Docket Number

01656.0011.PZUS00

ENCLOSURES (Check all that apply)

Fee Transmittal Form



Fee Attached



Amendment/Reply



After Final



Affidavits/declaration(s)



Extension of Time Request



Express Abandonment Request



Information Disclosure Statement



Certified Copy of Priority Document(s)

Reply to Missing Parts/
Incomplete ApplicationReply to Missing Parts
under 37 CFR 1.52 or 1.53

Drawing(s)



Licensing-related Papers



Petition

Petition to Convert to a
Provisional Application

Power of Attorney, Revocation



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Request for Refund



CD, Number of CD(s) _____



Landscape Table on CD



After Allowance Communication to TC

Appeal Communication to Board
of Appeals and InterferencesAppeal Communication to TC
(Appeal Notice, Brief, Reply Brief)

Proprietary Information



Status Letter

Other Enclosure(s) (please identify
below):Provisional Patent Application Cover Sheet (2
pgs.), and return-receipt postcard.

Remarks

SIGNATURE OF APPLICANT, ATTORNEY, OR AGENT

Firm Name

Howrey Simon Arnold & White, LLP

Signature

Printed name

Teddy C. Scott, Jr., Ph.D.

Date

November 8, 2004

Reg. No.

53,573

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Patent Application: **PTEN Inhibitors**
List of Contributors: Taxiarchis M. Georgiadis, Ph.D.,
Joseph Garlich, Ph.D.,
Xiaodong Peng, Ph.D.,
Jin Su, Ph.D.,
Tim C. Smith, Ph.D.,

Summary

We have developed an assay that measures PTEN activity and have tested compounds in search of potent PTEN inhibitors. To date we have tested approximately 250 compounds and have found some activity in four distinct chemical entities.

Background and Existing Knowledge- PTEN

Cellular processes are to some extent controlled by cycles of phosphorylation and dephosphorylation involving lipids and proteins. PTEN (phosphatase located on chromosome 10) is a dual specificity phosphatase which dephosphorylates an important lipid second messenger, phosphatidylinositol 3,4,5 phosphate [Ptlins(3,4,5)P3] to control cell division and apoptosis. It is mutated at high frequency in human malignant disease (incidence varies from 20% to 95% depending on tumor type). Preliminary data from the Durden group has implicated PTEN in the control of tumor-induced angiogenesis and the control of immunoreceptor signaling suggesting that this is a major drug target for control of angiogenesis and inflammatory signals.

What we hope to achieve

We believe an agent which would inhibit PTEN thereby augmenting levels of PIP3 would likely have therapeutic efficacy in a number of disease states associated with uncontrolled cell death and tissue damage

Therapeutic areas

Chemistry

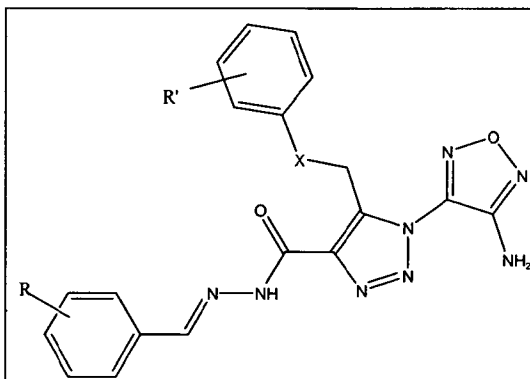
We have screened for compounds that would bind to PTEN and inhibit phosphorylation. PTEN hydrolyzes phosphate at the 3 position on the inositol ring of PtdIns(3,4,5)P3, and Ins(1,3,4,5)P4. The release of phosphate from the natural substrate was measured in a colorimetric assay by using the Malachite Green Reagent (Upstate) in accordance with the instructions of the manufacturer. The absorbance at 650 nm was recorded in an

ELISA plate reader. A standard curve was performed in each assay, and the amount of free phosphate was calculated from the standard curve line-fit data.

From an initial screening of 100 samples selected from insight using an *in silica* interaction of protein to commercially available samples, we were able to identify four initial series (and later a fifth series from literature screening) that shows some preliminary activity at 250uM concentrations. These series were pursued and we identified four distinct series that have been found to show modest activity in the PTEN assay. We have labeled the series as follows: 1) Furazan Series, 2) Diamide Series, 3) Sulf-hydrazone Series, and 4) Peptide Series. In addition, based on these results and searching relevant literature, a fifth series, the Diketone Series was discovered along with a number of other, miscellaneous compounds. All samples reported are represented using our internal numbering system (originally called “CC” numbers but later changed to our current “SF” numbering system) and individual lots are characterized by unique batch code numbers and notebook page numbers.

1.0 ChemNavigator Derived Series

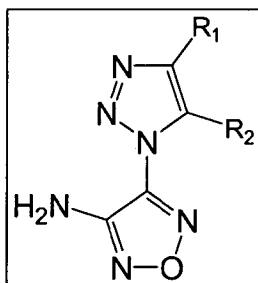
1.1 Furazan Series



| Batch Number | Compound Number | Notes | R Group (R'=H) | X Group | % Inhibition at 250uM | % Inhibition at 250uM | IC50 (uM) |
|--------------|-----------------|-----------|-----------------------|---------|-----------------------|-----------------------|-----------|
| BC100108 | CC1523-000 | 2nd batch | 3-(OEt),4-(OCH2CONH2) | O | 65 | 52.9 | 135 |
| BC100041 | CC1523-000 | | 3-(OEt),4-(OCH2CONH2) | O | 73.6 | 76.4 | |
| BC100033 | CC1515-000 | | 4-Br | N | 56 | | |
| BC100171 | CC1515-000 | | 4-Br | N | 38.3 | 46.2 | |

| | | | | | | | |
|----------|------------|--|-------|---|----|------|--|
| BC100143 | CC1623-001 | | 3-NO2 | O | 16 | 50.1 | |
|----------|------------|--|-------|---|----|------|--|

The initial hits **CC1523** and **CC1515** were explored. The core ring system was examined and a number of analogs were screened. As shown below, analogs that did not contain a phenyl hydrazide group in the R1 position and instead contained a precursor ethyl ester had little activity at 250uM. Likewise, it was apparent that aryloxy methylene or aryl amino methylene groups in the R2 position were important for activity.



| Compound Number | Barcode Number | Notebook Number | R1 | R2 | %Inhibition at 250uM | IC50 (uM) |
|-----------------|----------------|-----------------|--------------------------------------|-----------------|----------------------|-----------|
| CC1507-000 | BC100025 | A033-48 | CONHNCHPh(3Br) | CH2OMe | -14.25 | |
| CC1512-000 | BC100030 | A033-48 | CH3 | C(Me)NNH2 | 6.8 | |
| CC1515-000 | BC100033 | A033-47 | CONHNCHPh(4Br) | CH2NHPh | 56, 38.3 | |
| CC1515-000 | BC100171 | A033-70-22 | CONHNCHPh(4Br) | CH2NHPh | 46.2 | |
| CC1515-000 | BC100186 | A033-77-14 | CONHNCHPh(4Br) | CH2NHPh | 46.2 | |
| CC1521-000 | BC100039 | A033-47 | CONHNCHPh(3-OEt, 4-OCH2Ph(2-Cl,6-F)) | CH2NEt2 | -11.8 | |
| CC1523-000 | BC100108 | A033-56-9 | CONHNCHPh(3-OEt, 4-OCH2CONH2) | CH2OPh | 65, 52.8 | 245 |
| CC1523-000 | BC100041 | A033-47 | CONHNCHPh(3-OEt, 4-OCH2CONH2) | CH2OPh | 73.6, 76.4, 84.5 | 135 |
| CC1533-000 | BC100051 | A033-38-2 | CO2Et | CH2NHCHCH(OH)Ph | -20 | |
| CC1541-000 | BC100059 | A033-38-10 | CO2NHNCHPh(3-Br, 4-OH, 5-OMe) | CH2NEt2 | -10 | |
| CC1618-000 | BC100138 | A033-70-17 | CONHNHCOPh | Ph | -1.7 | |

| | | | | | | |
|------------|----------|------------|--|--|------------|--|
| CC1619-000 | BC100139 | A033-70-18 | CO ₂ NHNCHPh(3-Br, 4-OH, 5-OMe) | Ph | 18.7 | |
| CC1620-000 | BC100140 | A033-70-19 | CO ₂ NHNCHPh(3,4,5-triOMe) | Ph | 18.6 | |
| CC1623-001 | BC100143 | A033-70-23 | CONHNCHPh(3-NO ₂) | CH ₂ NHPh | 50.1 | |
| CC1623-001 | BC100187 | A033-77-15 | CONHNCHPh(3-NO ₂) | CH ₂ NHPh | 45.5 | |
| CC1633-000 | BC100153 | A033-88-1 | CO ₂ Et | CH ₂ NHPh | -11.1 | |
| CC1634-000 | BC100154 | A033-88-2 | CO ₂ Et | CH ₂ OPh | -14.4 | |
| CC1635-000 | BC100155 | A033-88-3 | CO ₂ Et | CH ₂ S-2benzo[d]thiazole | 21.2, 15.5 | |
| CC1636-000 | BC100156 | A033-88-4 | CO ₂ Et | CH ₂ -indoline | 9.2 | |
| CC1637-000 | BC100157 | A033-88-5 | CO ₂ Et | CH ₂ S-(1-Me-1H-imidazol) | -11.2 | |
| CC1638-000 | BC100158 | A033-88-6 | CO ₂ Et | CH ₂ N(Me)CH ₂ Ph | -4.1 | |
| CC1639-000 | BC100159 | A033-88-7 | CO ₂ Et | CH ₂ S(5-NH ₂ -1,3,4-thiadiazol-2-yl) | -9.1 | |
| CC1640-000 | BC100160 | A033-88-8 | CO ₂ Et | CH ₂ S(1H-benzo[d]imidazol-2-yl) | 2.1 | |
| CC1641-000 | BC100161 | A033-88-9 | CO ₂ Et | CH ₂ (1H-benzo[d][1,2,3]triazol-1-yl) | -7 | |
| CC1642-000 | BC100162 | A033-88-10 | CO ₂ Et | CH ₂ NHCH ₂ (2-Py) | 0.8 | |
| CC1643-000 | BC100163 | A033-88-11 | CO ₂ Et | CH ₂ S(4,6-dimethylpyrimidin-2-yl) | 0.3 | |
| CC1644-000 | BC100164 | A033-88-12 | CO ₂ Et | CH ₂ NHPh(4-OMe) | 12.4, 10.5 | |
| CC1645-000 | BC100165 | A033-88-13 | CO ₂ Et | CH ₂ N(CH ₂ CH ₂) ₂ N(2-Py) | 0.9 | |
| CC1726-000 | BC100262 | A048-41-1 | CO ₂ Et | NHCH ₂ Ph(3-Br, 4-Me) | 25.4 | |
| CC1728-000 | BC100264 | A048-41-3 | CONHNCHPh(3,4-diOMe) | Me | 34 | |

We plan to continue looking for additional analogs. With respect to the R1 Group, we plan to optimize the spacer unit (length and composition) as well as determine optimal substitution on the terminal aromatic ring and determine if substituted aliphatic rings

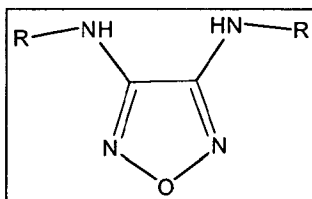
enhance activity). With respect to the R2 Group, we plan to optimize the spacer (currently -CH₂O-, -CH₂NH-, -NHCH₂-, -CH₂S-, -CH₂-) as well as determine optimal substitution on the terminal aromatic ring (and determine if substituted aliphatic rings enhance activity). We also plan to examine the significance of the amino furazan ring and determine the effect of substituting with other substituted aryl rings.

Diamide Series

The Diamide Series started out as a symmetrical molecule with a core ring system comprised of a furazan ring (**SF 1518**). Derivatives were synthesized to determine the inhibitory effect of the symmetrical R groups, the core ring system, and the symmetry of the molecules.

1.2.1 R Group Derivatives

1.2.2



From the initial study it appeared that **SF1609** and **SF1617** has an IC₅₀ ~100uM.

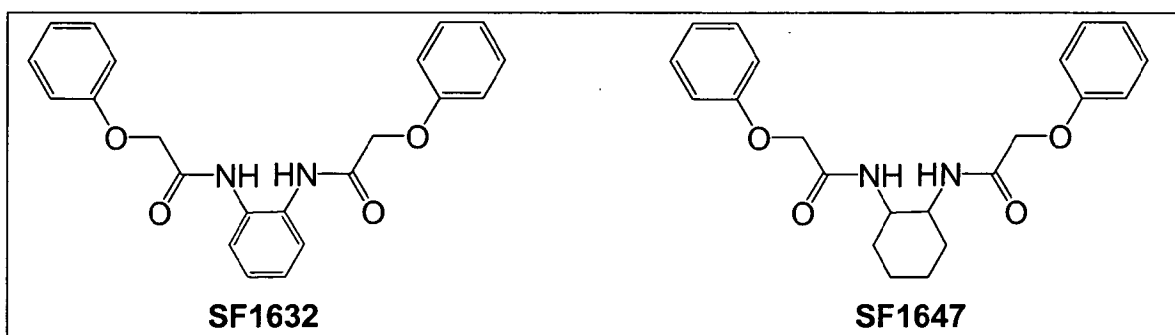
| SF Number | Notebook # | Barcode # | R | %Inh.@250uM | IC ₅₀ (uM) |
|------------|------------|-----------|-----------------------------------|----------------|-----------------------|
| SF1518-000 | A033-47 | BC100036 | COCH ₂ OPh(3-Me) | 46.6 | |
| SF1607-000 | A033-70-6 | BC100127 | COCH ₂ OPh(2-OMe) | 11.3 | |
| SF1607-000 | A033-77-2 | BC100174 | COCH ₂ OPh(2-OMe) | 10.7 | |
| SF1608-000 | A033-70-7 | BC100128 | COCH ₂ OPh(4-Br) | 41.8, 26.3 | |
| SF1608-000 | A033-77-3 | BC100175 | COCH ₂ OPh(4-Br) | 25.9 | |
| SF1609-000 | A033-70-8 | BC100129 | COCH ₂ OPh(2,5-diMe) | 77.7, 76, 79.8 | 80uM, 113uM |
| SF1609-000 | A033-77-4 | BC100176 | COCH ₂ OPh(2,5-diMe) | 58.4 * | |
| SF1610-000 | A033-70-9 | BC100130 | COCH ₂ OPh(2-iPr,5-Me) | 39.3 | |
| SF1610-000 | A033-77-5 | BC100177 | COCH ₂ OPh(2-iPr,5-Me) | 28.3 | |
| SF1611-000 | A033-70-10 | BC100131 | COCH ₂ OPh(4-OMe) | 33.8 | |
| SF1611-000 | A033-77-6 | BC100178 | COCH ₂ OPh(4-OMe) | 19.7 | |
| SF1612-000 | A033-70-11 | BC100132 | COCH ₂ OPh | 82 | |

| | | | | | |
|------------|------------|----------|-----------------|--------------|------|
| SF1612-000 | A033-77-7 | BC100179 | COCH2OPh | 3.3 | |
| SF1614-000 | A033-70-13 | BC100134 | COCH2OPh(2-Me) | 41.9 | |
| SF1614-000 | A033-77-9 | BC100181 | COCH2OPh(2-Me) | 56.5 | |
| SF1615-000 | A033-70-14 | BC100135 | COCH2OPh(4-Me) | 38.5 | |
| SF1615-000 | A033-77-10 | BC100182 | COCH2OPh(4-Me) | 46.2 | |
| SF1616-000 | A033-70-15 | BC100136 | COCH2OPh(2-iBu) | 13.5 | |
| SF1616-000 | A033-77-11 | BC100183 | COCH2OPh(2-iBu) | 40.8 | |
| SF1617-000 | A048-41-4 | BC100265 | COCH2OPh(2-iPr) | 75.5, -16.3* | 84uM |
| SF1617-000 | A048-43-4 | BC100270 | COCH2OPh(2-iPr) | 78.2, 3.4* | |
| SF1617-000 | A033-70-16 | BC100137 | COCH2OPh(2-iPr) | 81.1, 30.2* | 66uM |
| SF1617-000 | A033-77-12 | BC100184 | COCH2OPh(2-iPr) | 77.7, 34.7* | |
| SF1617-000 | A048-56-2 | BC100283 | COCH2OPh(2-iPr) | 39.6* | |

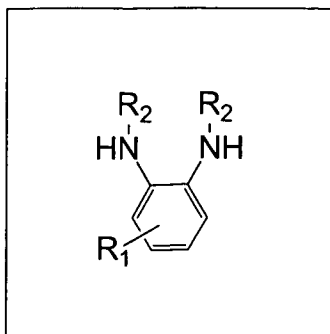
* Repeated using slightly different assay conditions.

1.2.2 Core Group Derivatives-Symmetrical Phenyl

In addition to modifying the terminal R groups we also examined the effect of substituting the furazan ring with other ring systems, both aromatic and aliphatic. We determined that a planar orientation was preferable based on the cyclohexane analog (**SF1647**, 2.2% inhibition@250uM) compared to the phenyl analog (**SF1632**, 31.3% inhibition@250uM). We plan to further examine the effect of 1,2 substitution with 1,3 and 1,4 substitution on the core aromatic ring.



We synthesized derivatives of **SF1647** and have shown the results in the table below. These compounds all have a 1,2 disubstituted aromatic core ring with symmetrical substitution.



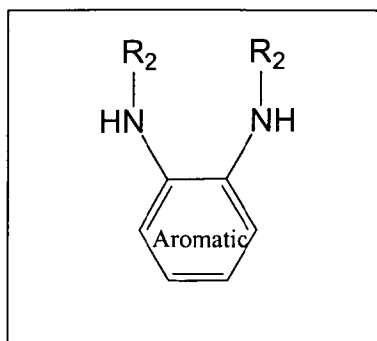
| SF Number | Notebook # | Barcode # | R1 | R2 | %Inh.@250uM |
|------------|------------|-----------|--------|-----------------|-------------------|
| SF1632-000 | A033-77-16 | BC100188 | H | COCH2OPh | 31.3 |
| SF1632-000 | A033-67D | BC100152 | H | COCH2OPh | 31.5 |
| SF1648-000 | A048-10-1 | BC100229 | H | COCH2OPh(2-iPr) | 4.9 |
| SF1648-000 | A033-84 | BC100168 | H | COCH2OPh(2-iPr) | -15.5 |
| SF1649-000 | A033-93 | BC100169 | H | COCH2OPh(2-OMe) | -13.2, -22.5 |
| SF1695-000 | A033-87-1 | BC100218 | H | COCH2OPh(4-OMe) | 34.4, 13.2* |
| SF1695-000 | A048-14B | BC100235 | H | COCH2OPh(4-OMe) | 40.2 |
| SF1701-000 | A033-92 | BC100223 | H | COCH2OPh(2-Me) | 51.4, 32.9 |
| SF1703-000 | A033-95T | BC100225 | H | COCH2OPh(3-OMe) | 32.8 |
| SF1712-000 | A048-19B | BC100237 | H | COCH2OPh(4-OMe) | 56.5, 69.3, 15.5* |
| SF1646-000 | A033-76-B | BC100167 | 4-Me | COCH2OPh | 33.7, 34.6, 1.8* |
| SF1696-000 | A033-76-C | BC100219 | 4-CO2H | COCH2OPh | 29.2 |
| SF1697-000 | A033-76-D | BC100220 | 3-Me | COCH2OPh | 9.1 |
| SF1713-000 | A048-20 | BC100238 | 4-Me | COCH2OPh(4-OMe) | 31.5, 66.5, 17.8* |
| SF1744-000 | A048-58-3 | BC100286 | 4CO2H | COCH2OPh(4-OMe) | -18 |

* Repeated using slightly different assay conditions.

We plan to continue looking for additional analogs. With respect to the R Group, we plan to optimize the spacer unit (length and composition) as well as determine optimal substitution on the terminal aromatic ring (and determine if substituted aliphatic rings enhance activity).

1.2.3 Core Group Derivatives-Symmetrical Aromatic Rings

In addition to phenyl rings, other aromatic rings were incorporated into the core ring system. These include pyridyl and pyrimidyl rings.



| SF Number | Notebook # | Barcode # | Center Ring | R2 | %Inh.@240uM |
|------------|------------|-----------|-------------|-----------------|-------------|
| SF1698-000 | A033-76-E | BC100221 | pyrimidine | COCH2OPh | 38.5, 36.1* |
| SF1747-000 | A048-58-6 | BC100289 | Pyridine# | COCH2OPh(4-OMe) | -33.5 |

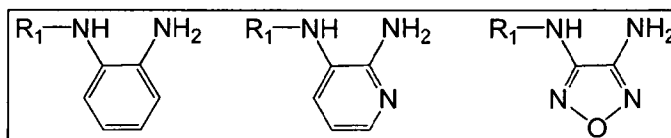
2,3 disubstituted pyridine

* Repeated using slightly different assay conditions.

We plan to examine the effect of replacing the furazan ring with a variety of other heterocycles, including pyridine, pyrimidine, thiophene, furan ring systems.

1.2.4 Core Group Derivatives-Asymmetrical Aromatic Rings

In addition, we examined the symmetrical nature of the series. We synthesized some mono-substituted core rings. Each compound contains only one side group on the aromatic di-amine.



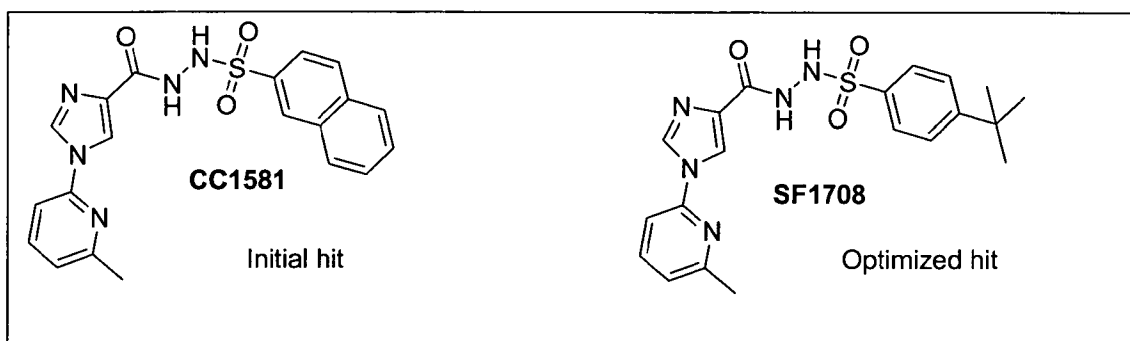
| SF Number | Notebook # | Barcode # | Core Ring | R1 | %Inh.@240uM | IC50 (uM) |
|------------|------------|-----------|----------------|-----------------|---------------|-----------|
| SF1700-000 | A033-96-A | BC100170 | benzene | COCH2OPh(2-iPr) | -18.4 | |
| SF1704-000 | A033-95B | BC100226 | benzene | COCH2OPh(3-OMe) | 56.6, (96.1)* | 220uM |
| SF1706-000 | A048-10-2 | BC100230 | benzene | COCH2OPh(2-iPr) | 10.6 | |
| SF1710-000 | A048-14A | BC100234 | benzene | COCH2OPh(4-Me) | 4.6 | |
| SF1711-000 | A048-19A | BC100236 | benzene | COCH2OPh(4-OMe) | -13.6 | |
| SF1742-000 | A048-58-1 | BC100284 | benzene (3-Me) | COCH2OPh(4-OMe) | -21.9 | |

| | | | | | | |
|------------|-----------|----------|-----------------|-----------------|-------|--|
| SF1745-000 | A048-58-4 | BC100287 | benzene(5CO2H) | COCH2OPh(4-OMe) | -25.4 | |
| SF1746-000 | A048-58-5 | BC100288 | pyridine(2-NH2) | COCH2OPh(4-OMe) | -3.5 | |
| SF1748-000 | A048-58-7 | BC100290 | furazan | COCH2OPh(4-OMe) | -11.9 | |

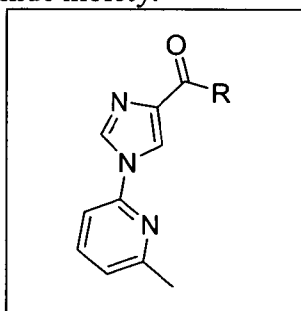
* Repeated using slightly different assay conditions.

We plan to continue looking for additional analogs. With respect to the symmetry, we plan to determine if unsymmetrical analogs display better activity. We plan to examine the role of the primary amine (in the above mentioned cases). Unsymmetrical disubstituted core ring systems are also envisioned.

1.3 Sulf-hydrazone Series



In the Sulf-hydrazone series, our initial hit **CC1581** was derivatized with emphasis on manipulating the terminal aryl amide moiety.



| Compound Number | Batch Code Number | R Group | % Inhibition at 250uM | IC50 (uM) |
|-----------------|-------------------|---------------------|-----------------------|-----------|
| SF1581-000 | BC100210 | NHNHSO2(2-Naphthyl) | 76.6 | 190 |
| SF1581-000 | BC100100 | NHNHSO2(2-Naphthyl) | 55.6, 41.3, (-3.5)* | |
| SF1688-000 | BC100211 | NHNH2 | 7.1 | |
| SF1689-000 | BC100212 | NHNHCOPh(4-Me) | 6 | |
| SF1690-000 | BC100213 | NHNHCOPh(4-Br) | 22.7 | |
| SF1691-000 | BC100214 | NHNHSO2Ph(4-OMe) | 6.9 | |

| | | | | |
|------------|----------|--|----------------------|----|
| SF1692-000 | BC100215 | NHNCHPh(4-NO ₂) | 45.5, 38.9, (-1.5)* | |
| SF1693-000 | BC100216 | NHNHSO ₂ Ph(3-CF ₃) | 13.6 | |
| SF1694-000 | BC100217 | NHNCHPh | 12.9 | |
| SF1699-000 | BC100222 | NHNHCO(2-naphthyl) | 42.8, 41.4, (-13.4)* | |
| SF1702-000 | BC100224 | NHNHSO ₂ Ph(4-Me) | 46.4, 43.7, (-10.1)* | |
| SF1707-000 | BC100231 | NHNHSO ₂ Ph(4-tBu) | 25.1 | |
| SF1708-000 | BC100268 | NHNHCOPh(4-tBu) | 95.5, 84.6 | 50 |
| SF1708-000 | BC100269 | NHNHCOPh(4-tBu) | -8.6* | |
| SF1708-000 | BC100232 | NHNHCOPh(4-tBu) | 56.9* | |
| SF1709-000 | BC100233 | NHNHCO(CH ₂) ₃ Ph | 28.5 | |
| SF1714-000 | BC100239 | OH | 8 | |
| SF1730-000 | BC100267 | NHNHCOPh(4-NO ₂) | -9.7 | |
| SF1731-000 | BC100271 | NHNHSO ₂ Ph(4-NO ₂) | 8.3* | |
| SF1739-000 | BC100280 | NHNCHPh(4-tBu) | -14.3* | |
| SF1775-000 | BC100319 | OMe | 18.3* | |

* Repeated using slightly different assay conditions.

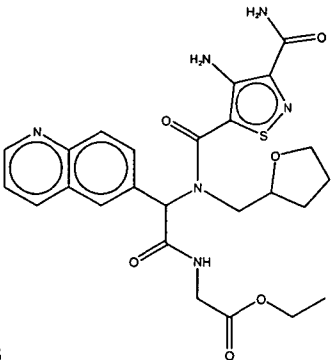
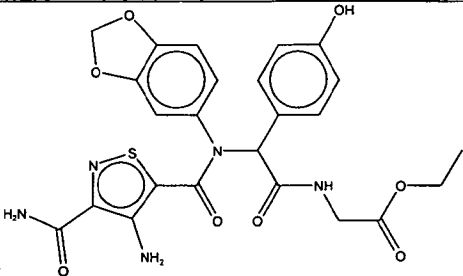
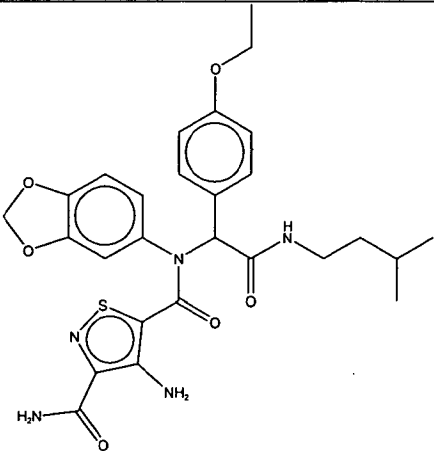
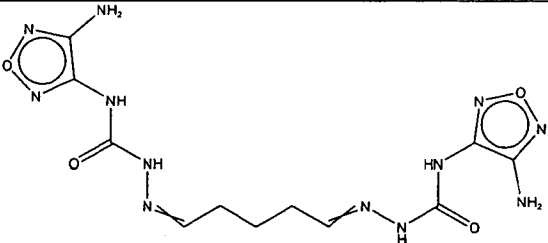
We plan to continue looking for additional analogs. We plan to examine the spacer unit between the biaryl and the terminal aryl ring. In addition we plan to incorporate other biaryl functionalities.

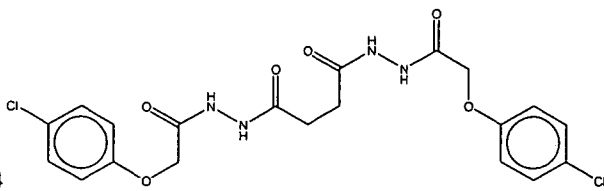
1.4 Peptide Series

1.5

From the initial screening of 100 compounds there were several hits that were peptide like. They consisted of long aliphatic chains with characteristic amide bonds. We have not optimized these series to date.

| Structure | Sample Number | Batch Number | Semafore Number | % Inhibiti on at 250uM |
|-----------|------------------|-----------------|--------------------|---------------------------------|
|-----------|------------------|-----------------|--------------------|---------------------------------|

| | | | | |
|--|------------|----------|------------|-------|
| <p>Structure86</p>  | | | | |
| | A033-55-12 | BC100085 | CC1566-000 | 70.6% |
| <p>Structure72</p>  | | | | |
| | A033-54-10 | BC100071 | CC1552-000 | 52.4% |
| <p>Structure80</p>  | | | | |
| | A033-55-6 | BC100079 | CC1560-000 | 37.1% |
| <p>Structure87</p>  | | | | |
| | A033-56-1 | BC100086 | CC1567-000 | 60.2% |

| | | | | |
|---|---------|----------|------------|------|
|  <p>Structure 34</p> | A033-47 | BC100034 | CC1516-000 | 35.3 |
|---|---------|----------|------------|------|

1.5 Summary of the ChemNavigator series

The four series initiated from the ChemNavigator *in silica* database search yielded several hits. For this specific effort, a hit is defined as a compound exhibiting greater than 30% inhibition at 250 uM in the PTEN assay. Our initial optimization of these series produced several micromolar inhibitors in the PTEN assay.

| SF# | PTEN | Series |
|--------|-------|-----------------------|
| SF1523 | 135uM | Furazan Series |
| SF1609 | 80uM | Diamide Series |
| SF1617 | 66uM | Diamide Series |
| SF1708 | 50uM | Sulf hydrazone Series |

2.0 Semafore Derived Series

Inspired from the work of Urbanek et al (*J.Med Chem* **2001**, *44*, 1777-1793) on their work on potent reversible Inhibitors of Protein Tyrosine Phosphatase CD45 and from our internal research efforts, we examined the PTEN activity of a series of diketone compounds. We hypothesize that the diketone moiety would be capable of reacting with the PTEN active site cystine and that we could engineer in selectivity by taking advantage of PTEN's larger catalytic site compared to other phosphatases. We have identified two series of compounds, labeled Diketone Phananthrolines and Diketone Phenanthrenes, that exhibit greater than 95% inhibition @ 250 uM in the PTEN assay. In addition, several miscellaneous compounds were found to have modest micromolar inhibition in the PTEN assay.

We examined a list of known PTP inhibitors and tested them in the PTEN assay. We discovered that several vanadate compounds (known PTP1B inhibitors) also exhibit PTEN activity. Recently the Woscholski group at Imperial College, England (Schmid *et al.*, *FEBS* **2004**, *566*, 35-38), reported PTEN activity for several oxovanadates (CC1668,

CC1664, CC1674, CC1675).

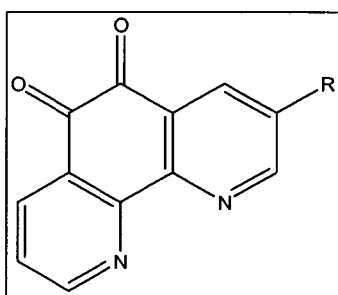
| Barcode Number | Sample Number | Compound Name | % Inhibition at 250 uM | IC50 (uM) |
|----------------|---------------|---|------------------------|-----------|
| BC100189 | CC1668-101 | Potassium Bisperoxo(bipyridine)oxovanadate (V) | 48.1 | |
| BC100190 | CC1669-100 | Alendronate, Sodium, Trihydrate | -0.9 | |
| BC100191 | CC1670-000 | N-(9,10-Dioxo-9,10-dihydro-phenanthren-2-yl)-2,2-dimethyl-propionamide | 99.7 | 2 |
| BC100192 | CC1671-000 | 5-Benzyl-3-furylmethyl (1R,S)-cis,trans-chrysanthemate | 9.6 | |
| BC100193 | CC1672-100 | Suramin, Sodium Salt; 8,8'-[carbonylbis[imino-3,1-phenylene carbonyl imino(4-methyl-3, 1-phenylene)carbonylimino]]bis-, hexa sodium salt | -9.9 | |
| BC100194 | CC1673-000 | 4-Methoxyphenacyl Bromide | 2 | |
| BC100195 | CC1674-101 | Dipotassium Bisperoxo(5-hydroxypyridine-2-carboxyl)oxovanadate (V) | 60 | |
| BC100196 | CC1675-101 | Dipotassium Bisperoxo(picolinato)oxovanadate (V) | 66.2 | |
| BC100197 | CC1676-000 | 1,4-Dimethylendothall; 1,4-Dimethyl-7-oxabicyclo[2.2.1]heptane-2,3-dicarboxylic Acid | 3.3 | |
| BC100198 | CC1677-000 | Monoperoxo(picolinato)oxovanadate(V) | 43.9 | |
| BC100199 | CC1678-101 | Potassium Bisperoxo(1,10-phenanthroline)oxovanadate (V) | 92 | 7 |
| BC100200 | CC1679-000 | Cantharidic Acid; 2,3-dimethyl-7-oxa-bicyclo[2.2.1]heptane-2,3-dicarboxylic acid | 2.7 | |
| BC100201 | CC1680-000 | Sodium Stibogluconate; Antimony Sodium Gluconate | 1.5 | |
| BC100202 | CC1681-000 | 3,4-Dephostatin, Ethyl- | 82.4 | 50 |
| BC100203 | CC1682-000 | bis(N,N-Dimethylhydroxamido)hydroxooxovanadate | 26.2 | |
| BC100204 | CC1683-000 | Fenvalerate; α -Cyano-3-phenoxybenzyl- α -(4-chlorophenyl)isovalerate | -25.5 | |
| BC100205 | CC1684-100 | α -Naphthyl Acid Phosphate, Monosodium Salt | 11.2 | |
| BC100206 | CC1685-100 | β -Glycerophosphate, Disodium Salt, Pentahydrate | 10.9 | |
| BC100207 | CC1686-000 | Endothall; 7-Oxabicyclo[2.2.1]heptane-2,3-dicarboxylic Acid | -1.2 | |
| BC100208 | CC1687-000 | Cypermethrin; (R,S)- α -Cyano-3-phenoxybenzyl-3-(2,2-dichlorovinyl)-2,2-dimethyl cyclopropanecarboxylate; (1R)-(R)-cyano(3-phenoxy phenyl) methyl 3-(2,2-dichloro vinyl)-2,2- dimethylcyclopropane carboxylate | -5.9 | |

| | | | |
|----------|------------|--|-----|
| BC100209 | CC1667-000 | Deltamethrin; (S)- α -Cyano-3-phenoxybenzyl(1R)-cis-3-(2,2-dibromovinyl)-2,2-dimethyl cyclopropanecarboxylate | 0.3 |
|----------|------------|--|-----|

2.0 Table of PTP Inhibitors tested in PTEN assay at 250uM. For select compounds, the IC50 value is recorded.

2.1 Diketone Phananthrolines

We are interested in exploring this series. **SF1720** (R=H) has low micromolar IC50. Attempts to derivatize the ring system are in progress.



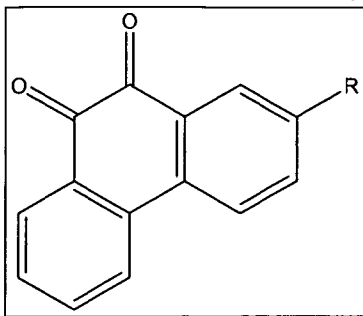
| Notebook Number | Barcode Number | SF Number | Reference | Substitution (R) | IC50 (nM) |
|-----------------|----------------|------------|-----------|------------------|-----------|
| A053-38 | BC100281 | SF1720-000 | A048-32-6 | H | 5000 |

We plan to optimize this series in a similar fashion to that of the Diketone Phenanthrenes. In addition, we feel that a non planer heterocyclic system (similar to benzyl), namely analogs of 1,2-di(pyridin-3-yl)ethane-1,2-dione, may have significant activity in the PTEN assay. We speculate that the pyridyl amines can coordinate with the protein via chelation to align the diketone functionality into the desired orientation producing an active series. Benzyl analogs, which lack coordination ability (**SF1729**) have little activity in the PTEN assay. We are pursuing the synthesis of 1,2-di(pyridin-3-yl)ethane-1,2-dione, and its analogs.

2.2 Diketone Phananthrenes

Another Diketone series that has activity in the PTEN assay is one where the diketone functionality is locked in a planer ring. Our first hit, **SF1670**, was quickly followed up by additional compounds designed to maximize interactions between the ligand and the protein. By adding aryl groups attached via a linker to the planer aromatic amine, we

were able to obtain nanomolar inhibitors of PTEN.



| Notebook Number | Barcode Number | Sample Number | Reference | Substitution | IC50 (nM) |
|-----------------|----------------|---------------|-----------|--|-----------|
| A033-90-3 | BC100191 | SF1670-000 | A033-90-3 | NHCOtBu | 2000 |
| A048-33-7 | BC100257 | SF1721-000 | A048-33-7 | H | 3400 |
| A048-33-8 | BC100258 | SF1722-000 | A048-33-8 | NO ₂ | 4000 |
| A053-38 | BC100281 | SF1740-000 | A053-28 | NHCOCH ₂ OPh | 400/ 327* |
| A048-58-10 | BC100293 | SF1751-000 | A048-57-1 | NHCOCH ₂ OPh(4-OMe) | 622.6 |
| A060-18 | BC100315 | SF1771-000 | A048-78-1 | NHCOCH ₂ OPh(4-Me) | 395.3 |
| A060-22 | BC100316 | SF1772-000 | A048-78-2 | NHCOCH ₂ OPh(2-iPr) | 435.6 |
| A060-60 | BC100317 | SF1773-000 | A048-78-3 | NHSO ₂ Ph | 221/ 297* |
| A060-74 | BC100318 | SF1774-000 | A048-78-4 | NHSO ₂ Ph(4-NO ₂) | 214.6 |
| A048-48 | BC100321 | SF1777-000 | A048-78-7 | NHCOPh | 291.7 |
| A048-73 | BC100323 | SF1779-000 | A048-78-9 | NHCOPh(4-Me) | 342.1 |
| A060-78 | BC100324 | SF1780-000 | A048-79-3 | NHSO ₂ Ph(4-tBu) | 269 |
| A060-92 | BC100328 | SF1784-000 | A048-83-1 | NHCOPh(2-NO ₂) | 598.7 |
| A060-96 | BC100329 | SF1785-000 | A048-83-2 | NHCO(CH ₂) ₃ Ph | 4998 |
| A060-98 | BC100330 | SF1786-000 | A048-83-3 | NHCOCO ₂ Et | 692.4 |
| A066-4B | BC100331 | SF1787-000 | A048-83-4 | NHCOCH ₂ OPh(2-OMe) | 548.1 |
| A066-6B | BC100332 | SF1788-000 | A048-83-5 | NHCOCH ₂ OPh(3-OMe) | 410.4 |
| A060-86 | BC100333 | SF1789-000 | A048-83-6 | NH ₂ | 776.1 |
| A066-02 | BC100334 | SF1790-000 | A048-83-7 | NHCOCH ₂ OPh(4-Cl) | 2620 |

* Run multiple times.

As seen in the above table, our best hits have additional functionality linked to the diketone ring lowering the inhibition into the nanomolar range. We are continuing to optimize this series by examining the length and constituency of the linker unit as well as

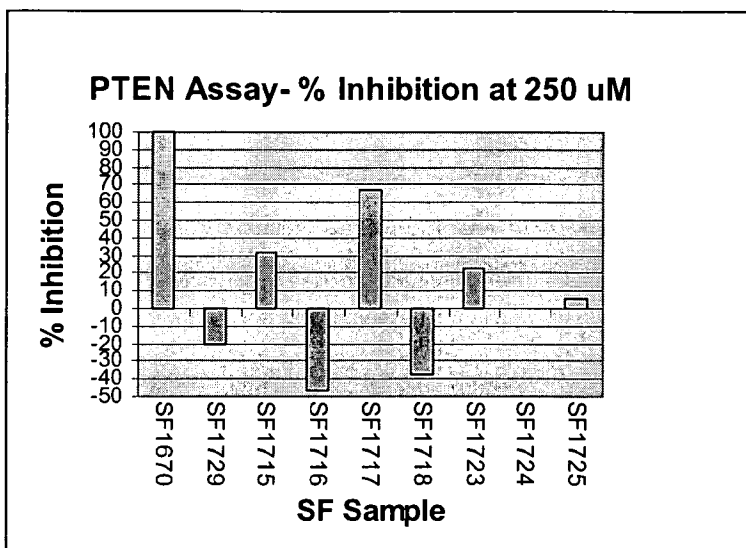
the choice of substituted aromatic ring on the end of the linker. In addition to aromatic rings, we are also planning aliphatic derivatives based on **SF1670**. We have examined the role of the diketone ring. It is our current belief that the diketone functionality needs to be locked in a planer orientation. As mentioned in the next section, a mono ketone in the phenanthrene ring system is also being explored (**CC1734**).

2.3 Diketone Discussion

One interesting compound, **CC1670** (later renamed **SF1670**), had an initial IC₅₀ in the single digit micromolar region. This compound was the first one in what we described as the Phenanthrene Diketone series. Researching this compound in the literature led us to several original papers describing how the compound was originally part of a library which exhibited CD45 inhibition. The researchers postulated that the diketone functionalities must be locked in a six member ring for suitable interactions with a CYS group in the active site. Several acyclic benzyl compounds, including benzyl (**SF1729**) were shown to exhibit little activity in the CD45 assay.

We decided to investigate compounds that resembled **SF1670** and **SF1729** and analogs in the PTEN assay. From the literature we were aware of the similarities of the active site in PTEN, CD45 as well as PTP1B. We were most interested in testing acyclic analogs of **SF1729** with the goal of obtaining selective inhibitors. We tested a series of acyclic analogs looking for activity in the PTEN assay, along with selectivity over CD45 and PTP1B.

In Graph 1, we have tabulated the results from the cyclic (**SF1670**) compound along with benzyl, the acyclic compound **SF1729**, and other acyclic analogs. Specifically the 4,4' di-Br, 3,3' di-MeO, 4,4' di-Me, 4,4' di-MeO, 4-OMe, 4-NH₂, and 4-CONH₂ analogs (**SF1715**, **SF1716**, **SF1717**, **SF1718**, **SF1723**, **SF1724**, **SF1725** respectively, Notebook page A048-32) were tested. The clear trend is that the acyclic compounds-as reported for the CD45 assay- did not show appreciable activity in the PTEN assay at 250 uM. In these compounds the two ketone functionalities are not restrained in a planer manner.



Graph 1: Initial Phenanthrene Diketone Series –Comparing Cyclic and Acyclic Analogs in the PTEN Assay at 250 uM

Researching structures similar to **SF 1670** we discovered a different series, which we labeled the Phenanthroline Diketone series. One of these, **SF1720** (Notebook page A048-32-6) showed similar activity as **SF1670** in the PTEN assay. The two structures are sterically similar however, are stereoelectronically very different. An analogy to visualize these distinct differences, can be to examine a simple benzene and pyridine ring-similar in shape but with profoundly different stereoelectronics. The Phenanthroline Diketone series has proven more difficult to derivatize, however, attempts are still in progress for further derivatives due to the preliminary selectivity results.

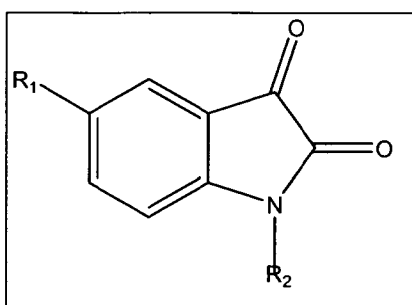
We have started optimizing the Phenanthrene Diketone series discovered in our labs. Starting from **SF1670** with an IC50 value of 2uM we have optimized this activity into the nanomolar range. We have manipulated the initial compound by inserting additional aromatic ring(s) attached via a linker. We have modified both the length and the structure of the linkers. We are currently attempting to optimize for selectivity with other phosphatases based on the crystal structure of the active site of PTEN protein.

3.0 Miscellaneous Dicarboxyl Compounds

3.1 Isatin Compounds

Based on the observed activity of the diketones, we investigated the Isatin Series (alpha

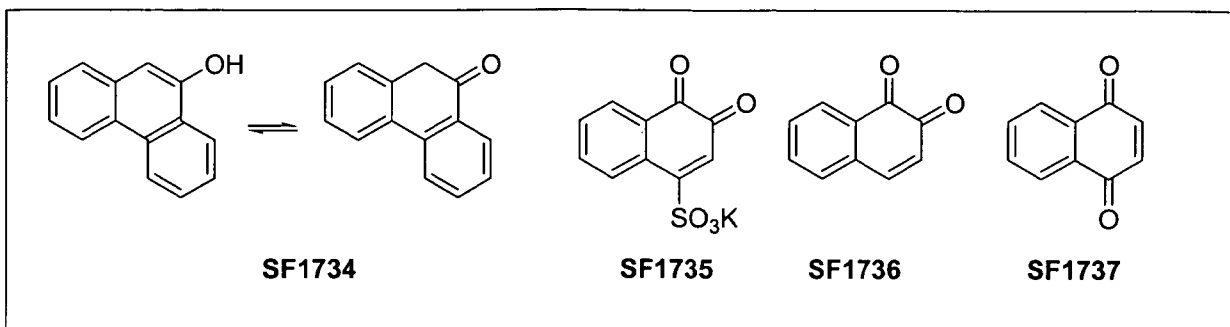
keto amides). These compounds have modest micromolar activity in the PTEN assay. We have just started SAR work on the series and plan to add functionality in three specific areas on the template molecule. Namely, attaching exploitable functionality on the aromatic ring, derivitizing via the amide NH group, and reacting the cyclic ketone to yield imine or olefinic derivatives. We have found that manipulation of 5-nitroindoline-2,3-dione (SF1770) by reducing the nitro group on the aromatic ring and attaching aryl groups via amide linkers, and by adding additional functionality on the amide nitrogen, can effect the inhibition of these compounds in the PTEN assay. The initial compounds synthesized and purchased exhibit low micromolar activity. The IC₅₀ for our two best compounds to date (SF1770 and SF1773) are in the double digit micromolar range.



| notebook_number | barcode_Number | SF Number | R1 | R2 | IC50 (uM) |
|-----------------|----------------|------------|-----------------------------|------------------------------|-----------|
| A048-66-20 | BC100313 | SF1770-000 | 5-NO ₂ | H | 2.2 |
| A048-70 | BC100322 | SF1778-000 | 5-(NHCOPh(4-Me)) | H | 22.05 |
| A048-77 | BC100325 | SF1781-000 | 5-(NHCOCH ₂ OPh) | H | 18.9 |
| A048-79-2 | BC100327 | SF1783-000 | 5-NO ₂ | CH ₂ Ph(2,4-diCl) | 4.6 |
| A048-83-8 | BC100335 | SF1791-000 | 5-H | CH ₂ Ph(4-Me) | >250 |
| A048-83-9 | BC100336 | SF1792-000 | 5-H | SO ₂ Ph(4-F) | >250 |
| A048-83-10 | BC100337 | SF1793-000 | 5-Me | CH ₂ Ph(4-Cl) | >250 |
| A048-83-11 | BC100338 | SF1794-000 | 5-iPr | H | >250 |
| A048-83-12 | BC100339 | SF1795-000 | 5-Br | Et | 100.459 |

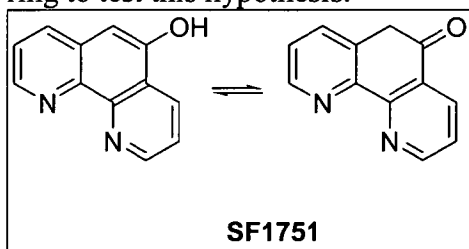
3.2 Miscellaneous Diketone Compounds

To determine what role the diketones played in PTEN activity, we have begun to examine a variety of dicarbonyl compounds and have discovered significant PTEN activity at 250uM. These compounds all exhibited low double digit micromolar IC₅₀ activity in the PTEN assay.



| Compound | PTEN Ic50 |
|----------|-----------|
| CC1734 | 33uM |
| CC1735 | 20uM |
| CC1736 | 21uM |
| CC1737 | 43uM |

We plan to derivatize the **SF1734** series in a similar fashion to that described for the Diketone series. In addition, we plan to functionalize the center ring system. One interesting observation has been the comparison of PTEN activity of **SF1734** and **SF1751**. It appears the heterocyclic compound as little activity at 250uM while the all carbon ring system (**SF1734**) has an IC50 value of 33uM. It could be that the stereoelectronics of the heterocyclic ring system is pushing the equilibrium to the enol form and thus reducing activity in the PTEN assay. We plan to make analogs of **SF1734** which contain both electron withdrawing and electron donating substitution on the core ring to test this hypothesis.



Because PTEN acts against PI3 kinase we believe that PTEN inhibitors will allow for efficient PI3 kinase activity in cells which will impart beneficial characteristics as hypothesized herein.

4.0 PTEN Use/ Utility

- PTEN inhibitors may protect against septic shock. The following article supports this idea. The authors summarize that their work suggests that stimulation of the PI3K pathway may be an effective approach for preventing or treating sepsis and/or septic shock.

Williams, David L.; Li, Chuanfu; Ha, Tuanzhu; Ozment-Skelton, Tammy; Kalbfleisch, John H.; Preiszner, Johanna; Brooks, Lynne; Breuel, Kevin; Schweitzer, John B. Modulation of the phosphoinositide 3-kinase pathway alters innate resistance to polymicrobial sepsis *Journal of Immunology* **2004**, 172(1), 449-456

ABSTRACT

We examined the effect of modulating phosphoinositide 3-kinase (PI3K) activity in a murine model of cecal ligation and puncture-induced polymicrobial sepsis. Inhibition of PI3K activity with wortmannin increased serum cytokine levels and decreased survival time in septic mice. We have reported that an immunomodulator, glucan phosphate, induces protection in murine polymicrobial sepsis. We observed that glucan stimulated tissue PI3K activity, which positively correlated with increased survival in septic mice. We investigated the effect of PI3K inhibition on survival in septic mice treated with glucan. Treatment of mice with the PI3K inhibitors, wortmannin and LY294002, completely eliminated the protective effect of glucan, indicating that protection against septic mortality was mediated through PI3K. Inhibition of PI3K resulted in increased serum levels of IL-1 β , IL-2, IL-6, IL-10, IL-12, and TNF- α in septic mice. Apoptosis is thought to play a central role in the response to septic injury. We observed that inhibition of PI3K activity in septic mice resulted in increased splenocyte apoptosis and a change in the anatomic distribution of splenocyte apoptosis. We conclude that PI3K is a compensatory mechanism that suppresses proinflammatory and apoptotic processes in response to sepsis and/or inflammatory injury. Thus, PI3K may play a pivotal role in the maintenance of homeostasis and the integrity of the immune response during sepsis. We also observed that glucan phosphate decreased septic morbidity and mortality through a PI3K-dependent mechanism. This suggests that stimulation of the PI3K pathway may be an effective approach for preventing or treating sepsis and/or septic shock.

-
- PTEN inhibitors may be useful in therapeutic angiogenesis. Sustained release of PTEN inhibitors (via nanoparticle technology or other slow in vivo release) may lead to therapeutic angiogenesis. See following article.

Yeh, J. L., Giordano, F. J., Gene-based therapeutic angiogenesis. *Semin Thorac Cardiovasc Surg* **2003**, 15, 236-249

ABSTRACT

Stimulating new blood vessel growth in ischemic hearts or limbs is a

hopeful new approach for patients with advanced vascular disease. This approach is based generally upon the hypothesis that sufficient exposure of a vascular bed to an >angiogenic< protein will stimulate neovascularization. Most >angiogenic< proteins have a markedly short serum half-life. To overcome this, researchers **have turned to gene therapy to ensure continuous expression of >angiogenic< proteins and prolonged exposure in the targeted vascular beds**. This field is still evolving, and although early >clinical< >trial< results suggest >angiogenic< gene therapy can be successful, many questions remain. As we continue to learn more about the complex interplay and coordinated action of the various factors involved in regulating >angiogenesis<, it is likely that strategies for therapeutic >angiogenesis< will continue to change. This review addresses the current state of >angiogenic< gene therapy, contrasts gene therapy with >angiogenic< protein delivery, describes early and recent >clinical< >trial< data, and discusses potential new directions in the field.;

More therapeutic angiogenesis (ie possible use for PTEN inhibitors):

Kleiman, Neal S.; Patel, Nirav C.; Allen, Keith B.; Simons, Michael; Yla-Herttuala, Seppo; Griffin, Elaine; Dzau, Victor J. Evolving revascularization approaches for myocardial ischemia. *American Journal of Cardiology* 2003, 92(9B), 9N-17N

ABSTRACT

Stable angina pectoris secondary to ischemic heart disease is a common and disabling condition. Medical therapy aims to relieve symptoms, improve exercise capacity, and decrease cardiac events by reducing myocardial oxygen demand or improving coronary blood supply to the ischemic myocardium. If medical treatment is inadequate, invasive revascularization procedures to improve coronary perfusion are considered. Percutaneous transluminal coronary angioplasty (PTCA) and coronary artery bypass graft (CABG) surgery are well-established and widely used myocardial revascularization techniques. Recent advances in PTCA have attempted to address the problem of restenosis, initially through the deployment of bare metal intracoronary stents and, more recently, with drug-eluting stents. Developments in CABG have focused on reducing the invasiveness of the procedure and minimizing the incidence of serious complications. Refinements include the use of mechanical stabilizers, endoscopic harvesting of conduit vessels, robotic telemanipulation systems, and fully automated anastomotic devices. Surgical laser transmyocardial revascularization and therapeutic >angiogenesis< represent newer approaches to coronary revascularization. Therapeutic >angiogenesis< aims to deliver an >angiogenic< growth factor or cytokine to the myocardium to stimulate collateral blood vessel growth throughout the ischemic tissue. The >angiogenic< factor may be administered as a recombinant protein or as a transgene within a plasmid or gene-transfer vector. Ongoing >angiogenic< gene therapy >clinical< >trials< are evaluating which factors, vectors, and delivery techniques hold the greatest promise for management of patients with chronic stable angina.

- Antisense PTEN inhibitor for diabetic use.

<http://www.genetrove.com/inVivoGT.html>

These data demonstrate that PTEN antisense will sensitize tissues to insulin resulting in decreased blood glucose concentrations in diabetic mice, but will not affect blood glucose levels in normal mice.

-
- Protection of cells is not guaranteed if p53 is inhibited.

Bonini, P.; Cicconi, S.; Cardinale, A.; Vitale, C.; Serafino, A. L.; Ciotti, M. T.; Marlier, L. N. Oxidative stress induces p53-mediated apoptosis in glia: p53 transcription-independent way to die. *J Neurosci. i Res.* **2004** (75), 83-95

ABSTRACT

Oxidative stress has been implicated in the pathogenesis of stroke, traumatic brain injuries, and neurodegenerative diseases affecting both neuronal and glial cells in the central nervous system (CNS). The tumor suppressor protein p53 plays a pivotal function in neuronal apoptosis triggered by oxidative stress. We investigated the role of p53 and related molecular mechanisms that support oxidative stress-induced apoptosis in glia. For this purpose, we exposed C6 glioma cells and primary cultures of rat cortical astrocytes to an H₂O₂-induced oxidative stress protocol followed by a recovery period. We evaluated the effects of >pifithrin-alpha< (PF-alpha), which has been reported to protect neurons from ischemic insult by specifically inhibiting p53 DNA-binding activity. Strikingly, PF-alpha was unable to prevent oxidative stress-induced astrocyte apoptosis. We demonstrate that p53 is able to mediate an apoptotic response by direct signaling at mitochondria, despite its transcriptional activity. The z-VAD-fmk-sensitive apoptotic response requires a caspase-dependent MDM-2 degradation, leading to p53 mitochondrial targeting accompanied by cytochrome c release and nucleosomal fragmentation.;

-
- Combination steroid treatment or inhibition in combo with PTEN inhibitors. (Endometreosis market for PTEN agonisits?) See following article.

Guzeloglu-Kayisli, Ozlem; Kayisli, Umit A.; Al-Rejjal, Rafat; Zheng, Wenxin; Luleci, Guven; Arici, Aydin Regulation of PTEN (phosphatase and tensin homolog deleted on chromosome10) expression by estradiol and progesterone in human endometrium . *Journal of Clinical Endocrinology & Metabolism* **2003**, 88(10), 5017-5026

ABSTRACT

PTEN (phosphatase and tensin homolog deleted on chromosome 10) is a

>tumor< suppressor gene, mutated frequently in a variety of human >tumors<. PTEN regulates cell growth, apoptosis, and proliferation. Phosphorylation in PTEN tail causes its inactivation and decreases its degradation. There is little known about the regulation of PTEN by ovarian steroids. We hypothesized that PTEN expression in human endometrium is variable throughout the menstrual cycle and early pregnancy, and that ovarian steroids regulate PTEN expression because PTEN is critical in many steroid-sensitive tissues such as endometrium, prostate, and breast. In the present study, we have observed a direct regulation of PTEN by ovarian steroids. Estradiol increased PTEN phosphorylation at 5-15 min. After 24-h treatment, progesterone induced a significant increase in PTEN protein levels, assessed by Western blot. Furthermore, we evaluated for the first time a comparison between menstrual cycle and early pregnancy, immunohistochemically. Endometrial PTEN expression revealed temporal and spatial changes throughout the menstrual cycle and during early pregnancy. We conclude that estradiol may downregulate PTEN activity by increasing its phosphorylation, but progesterone is likely to regulate the PTEN pool by decreasing its phosphorylation and increasing its protein level. Presented data, therefore, suggest that ovarian steroids regulate the endometrial PTEN pool. We propose that PTEN might be one of the signaling proteins that estrogen and progesterone are acting to affect endometrial cell proliferation and/or apoptosis.

- Possible use as notch ligand as targeting agent for PTEN inhibitors

Jundt, Franziska; Proebsting, Kristina Schulze; Anagnostopoulos, Ioannis; Mathas, Stephan; Stein, Harald; Doerken, Bernd Activated Notch signaling might be a novel therapeutic target for multiple myeloma. *Blood* 2003, 11, 928a

ABSTRACT

Notch signaling plays a key role in the development and differentiation of various hematopoietic lineages. In the hematopoietic system, Notch receptors are expressed in early hematopoietic stem cells, whereas Notch ligands are found in bone marrow stroma, which provides the microenvironment necessary for stem cell survival and differentiation. In addition, we recently demonstrated that Notch signaling is involved in the pathogenesis of B-cell-derived tumor cells of Hodgkin lymphoma (*Blood*. 2002;99:3398-3403). We described a novel mechanism for the oncogenic capacity of Notch by showing that interactions of overexpressed intact Notch1 and Notch2 receptors on tumor cells with their cognate ligand Jagged1 dramatically induce both proliferation and inhibition of apoptosis in vitro. We further provided evidence that in Hodgkin lymphoma Jagged1 is expressed in malignant as well as in bystander cells co-localizing with Notch-positive tumor cells. Notch signaling may therefore be activated in tumor cells by Jagged1 through homotypic or heterotypic cell-cell interactions and it seems likely that these interactions also contribute to lymphomagenesis in vivo. However, a pathogenetic role for Notch in multiple myeloma (MM), where tight interactions between neoplastic plasma cells and their microenvironment are essential for tumor cell growth, is currently unknown. In this study, we therefore investigated Notch gene expression in cultured and primary multiple myeloma cells. To that end, we analyzed 14 cases of MM for expression of Notch1 and Notch2 by immunohistochemistry. In all cases Notch1 and Notch2 were highly expressed

in MM cells. Strong Notch expression in MM cells was comparable to tumor cells of classic Hodgkin lymphoma, that we analyzed in our recent study. In contrast, we found low to undetectable levels of Notch1 and Notch2 in plasma cells of bone marrow of normal donors and in plasma cells of reactive lymphoid tissue. To verify high expression of Notch1 and Notch2 in cultured MM cells, we performed Western blot analysis of five MM cell lines. According to our data in primary MM cells, we found that both Notch receptors were highly expressed in all MM cell lines. However, freshly isolated CD19+ B cells and CD19+ B cells, that we differentiated to CD38+ plasmablastic cells in vitro, were almost completely devoid of Notch expression. Our data indicate that cultured and primary MM cells differ from their non-neoplastic counterparts with respect to strong Notch1 and Notch2 expression. Our data further provide evidence that **ligand-induced Notch signaling is a novel growth factor for multiple myeloma cells and suggest that these interactions contribute to lymphomagenesis of multiple myeloma in vivo**. Studies are under way to block Notch signaling by gamma-secretase inhibitors to further determine its role in tumor cell proliferation and resistance towards apoptosis in MM.

- PTEN inhibitors could help neural stem cell self-renewal

Erickson, R. I.; Groszer, M.; Ngo, C.; Liu, X.; Wu, H.; Kornblum, H.I. Serial passages of cortical neurospheres from conditional Pten mutant mice demonstrate persistent effects on neural stem cell self - renewal.- Society for Neuroscience Abstract Viewer and Itinerary Planner VOL. 2003 2003 PP. Abstract No. 243.18

ABSTRACT

Pten is a lipid phosphatase that acts as a >tumor< suppressor gene and is frequently mutated in gliomas. As one of its primary actions, Pten inhibits the PI3K/Akt pathway; therefore cells that have lost Pten function are enlarged, have enhanced survival, and increased proliferative capacity. Conditional knockout of Pten in nestin-containing cells results in enlarged, disorganized brains, and increased BrdU labeling in ventricular zones (Groszer et al. 2001). Using the neurosphere (NS) culturing system, we showed that Pten mutant (MUT) cells had increased proliferation mediated by more rapid cycling, as well decreased apoptosis. Neural stem cells (NSC) were more capable of self-renewal for at least one passage, but it **is not known whether this effect persists through multiple passages**. In the current study, we examine the effects of Pten deletion on multiply passaged neurospheres to determine whether effects on proliferation and self-renewal persist over multiple passages and whether there is an effect on cell fate specification. Cells from E14 MUT and wildtype (WT) cortices were plated at 5000 cells per ml and passaged several times. At each passage, aliquots of NS from each individual embryo were counted, measured, differentiated, and immunostained for neurons, astrocytes and oligodendrocytes. The number of NS generated remained higher in MUT over many passages, while mutant diameter measurements merged closer to WT. **The percent of tri-potential NS as well as the ratio of neurons/total cell counts remained higher in MUT**. This suggests that, **although proliferation slows in MUT, there are more NSC going through self-renewing divisions over subsequent passages**

More on PTEN and stem cells

Groszer, M.; Erickson, R.; Scripture-Adams, D. D.; Lesche, R.; Trumpp, A.; Zack, J. A.; Kornblum, H. I.; Liu, X.; Wu, H. Negative regulation of neural stem/progenitor cell proliferation by the Pten tumor suppressor gene in vivo. *Science*, VOL. 294 NO. 5549 2001 Dec 7 PP. 2186-9

ABSTRACT

The mechanisms controlling neural stem cell proliferation are poorly understood. Here we demonstrate that the PTEN >tumor< suppressor plays an important role in regulating neural stem/progenitor cells in vivo and in vitro. Mice lacking PTEN exhibited enlarged, histoarchitecturally abnormal brains, which resulted from increased cell proliferation, decreased cell death, and enlarged cell size. Neurosphere cultures revealed a greater proliferation capacity for tripotent Pten^{-/-} central nervous system stem/progenitor cells, which can be attributed, at least in part, to a shortened cell cycle. However, cell fate commitments of the progenitors were largely undisturbed. Our results suggest that PTEN negatively regulates neural stem cell proliferation.

- PTEN inhibitors for preventing neurodegenerative disease

Waldmeier, Peter C.; Tatton, William C., Interrupting apoptosis in neurodegenerative disease: Potential for effective therapy? *Drug Discovery Today* VOL. 9 NO. 5 1 March, 2004 PP.210-218

ABSTRACT

Current treatment options for neurodegenerative diseases are limited and mainly affect only the symptoms of disease. Because of the unknown and probably multiple causes of these diseases, they cannot be readily targeted. However, it has been established that apoptosis contributes to neuronal loss in most neurodegenerative diseases. A possible treatment option is to interrupt the signaling networks that link neuronal damage to apoptotic degradation in neurodegeneration. The viability of this option depends upon the extent to which apoptosis accounts for neuron loss, whether or not interruption of apoptosis signaling results in recovery of neurological function and whether or not there are significant downsides to targeting apoptosis. Several compounds acting at different sites in known apoptotic signaling networks are currently in development and a few are in clinical trial. **If an apoptosis-targeted compound succeeds in slowing or halting neurological dysfunction in one or more neurodegenerative diseases, a new era in the treatment of neurodegenerative diseases will begin.**

- Possible use for the prevention of apoptosis in mature erythroid progenitor cells.

Here is some important info; by inference a PTEN inhibitor would act opposite the effects noted here for the LY compound on erythroid progenitor

Paiboonsukwong, K.; Choi, I.; Matsushima, T.; Abe, Y.; Nishimura, J.; Winichagoon, P.; Fucharoen, S.; Nawata, H.; Muta, K. The signaling pathways of erythropoietin and

interferon-gamma differ in preventing the apoptosis of mature erythroid progenitor cells.
Int J Hematol VOL. 78 2003 Dec PP. 421-8

Interferon (IFN)-gamma is a survival factor for mature erythroid progenitor cells. To elucidate related survival mechanisms, we compared the role of phosphatidylinositol 3-kinase (PI3-kinase) in the survival signals of IFN-gamma and erythropoietin (EPO). Human erythroid colony-forming cells (ECFCs) purified from peripheral blood were used, and >Ly294002< was used as a PI3-kinase inhibitor. Treating ECFCs with a high concentration of >Ly294002< (50 micromol/L) in the presence of EPO and/or IFN-gamma reduced cell viability by inducing apoptosis. However, treating cells with a lower concentration of >Ly294002< (10 micromol/L) did not affect the antiapoptotic function of IFN-gamma and abolished the antiapoptotic effect of EPO. Adding IFN-gamma or EPO induced Bcl-x expression in ECFCs, as determined by Western blotting, and expression was suppressed in the presence of >Ly294002<. We also examined the phosphorylation of the protein kinase Akt, the downstream target of PI3-kinase. EPO stimulation significantly increased the level of Akt phosphorylation, but IFN-gamma did not. These results suggest that IFN-gamma plays a role in preventing the apoptosis of erythroid progenitor cells by affecting Bcl-x expression, thereby reducing the disruption of the mitochondrial transmembrane potential via PI3-kinase pathways that are related to but distinct from the EPO pathway

-
- Possible anti obesity therapeutic area. We expect our PTEN inhibitors would increase the effect of leptin in vivo by activating the PI3 kinase pathway

Huang, Wan; Dedousis, Nikolas; Bhatt, Bankim A.; O'Doherty, Robert M., Impaired activation of phosphatidylinositol 3-kinase by leptin is a novel mechanism of hepatic leptin resistance in diet-induced obesity - Journal of Biological Chemistry VOL. 279 NO. 21 May 21, 2004 PP. 21695-21700

ABSTRACT

Obesity is associated with the development of leptin resistance. However, the effects of leptin resistance on leptin-regulated metabolic processes and the biochemical defects that cause leptin resistance are poorly understood. We have addressed in rats the effect of diet-induced obesity (DIO), a situation of elevated tissue lipid levels, on the well described lipid-lowering effect of leptin in liver, an action that is proposed to be important for the prevention of tissue lipotoxicity and insulin resistance. In addition, we have addressed the role of phosphatidylinositol 3-kinase (PI 3-kinase) in mediating the acute effects of leptin on hepatic lipid levels in lean and DIO animals. A 90-min leptin (dollar sign 10 ng/ ml) perfusion of isolated livers from lean animals decreased triglyceride levels by 42 +/- 5% (p = 0.006). However, leptin concentrations ranging from dollar sign 10 to dollar sign 90 ng/ml had no effect on triglyceride levels in livers from DIO animals. The acute lipid-lowering effect of leptin on livers from lean animals was mediated by a PI 3- kinase-dependent mechanism, **because wortmannin and >LY294002<, the PI 3- kinase inhibitors, blocked the effects of leptin on hepatic**

triglyceride levels and leptin increased liver PI 3-kinase activity by 183 +/- 6% (p = 0.003) and insulin receptor substrate 1 tyrosine phosphorylation by 185 +/- 30% (p = 0.02) in the absence of PI 3-kinase inhibitors. Contrary to the effects of leptin in lean livers, leptin did not activate PI 3-kinase in livers from DIO rats. These data present evidence for a role for 1) leptin resistance in contributing to the excessive accumulation of tissue lipid in obesity, 2) PI 3-kinase in mediating the acute lipid-lowering effects of leptin in liver, and 3) defective leptin activation of PI 3-kinase as a novel mechanism of leptin resistance.

- PTEN inhibitors may block detrimental caspase-3 damage. Useful in diabetes and kidney failure.

Gao, Yong Mei; Debigare, Richard; Meireles, Christiane; Bailey, James

L; Price, S. Russ Insulin suppresses caspase-3-mediated actin cleavage and muscle proteolysis in L6 muscle cells: Implications for muscle atrophy-FASEB Journal VOL. 18 NO. 4-5 2004 PP. Abst. 827.8.

Muscle atrophy in catabolic conditions (e.g., diabetes, kidney failure) results from accelerated protein degradation (PD) by the ubiquitin-proteasome (Ub-P) system. Insulin resistance may be a signal for increased PD because: 1) insulin suppresses PD in muscle; and 2) insulin insufficiency stimulates muscle PD. We found that caspase-3 cleaves actin into fragments that are degraded by the Ub-P system in muscle. To examine the relationships between insulin, proteolysis, caspase-3 and actin cleavage in muscle, L6 muscle cells were incubated in medium with 0.5% serum +/- insulin as a model of insulin insufficiency. Serum deprivation (SD) increased PD 17% ($P < 0.05$), induced caspase-3 activity and increased the cleavage actin. Insulin attenuated these responses but the inhibitory effects required treatment for >4 h. In SD cells, insulin did not reduce the amount of procaspase-3 protein but increased several inhibitors of apoptosis (IAP) proteins which interact with caspase-3. Addition of >LY294002<, an inhibitor of PI 3-kinase, partially blocked the insulin-induced suppression of PD, and actin cleavage. In muscle of rats with kidney failure or acute diabetes, caspase-3 activity was higher and actin fragments were more abundant than in pair-fed, control rats. These data indicate that **insulin acts through the PI 3-kinase and other pathways to regulate caspase-3 activity and PD in muscle.** Our findings also suggest that insulin resistance is a stimulus for proteolysis in muscle.

- Inhibition of PTEN thus should enhance PDGF response and promote angiogenesis and other PDGF related activities.

Mahimainathan, L.; Choudhury, G. G. Inactivation of platelet-derived growth factor receptor by the tumor suppressor PTEN provides a novel mechanism of action of the phosphatase. -J Biol Chem VOL. 279 NO. 15 2004 Apr 9 PP. 15258-68

ABSTRACT

PTEN, mutated in a variety of human >cancers<, is a dual specificity protein phosphatase and also possesses D3-phosphoinositide phosphatase activity on phosphatidylinositol 3,4,5-tris-phosphate (PIP(3)), a product of phosphatidylinositol 3-kinase. This PIP(3) phosphatase activity of PTEN contributes to its >tumor< suppressor function by inhibition of Akt kinase, a direct target of PIP(3). We have recently shown that Akt regulates PDGF-induced DNA synthesis in mesangial cells. In this study, we demonstrate that expression of PTEN in mesangial cells inhibits PDGF-induced Akt activation leading to reduction in PDGF-induced DNA synthesis. As a potential mechanism, we show that PTEN inhibits PDGF-induced protein tyrosine phosphorylation with concomitant dephosphorylation and inactivation of tyrosine phosphorylated and activated PDGF receptor. Recombinant as well as immunopurified PTEN dephosphorylates autophosphorylated PDGF receptor in vitro. Expression of phosphatase deficient mutant of PTEN does not dephosphorylate PDGF-induced tyrosine phosphorylated PDGF receptor. **Rather its expression increases tyrosine phosphorylation of PDGF receptor. Furthermore, expression of PTEN attenuated PDGF-induced signal transduction including phosphatidylinositol 3-kinase and Erk1/2 MAPK activities. Our data provide the first evidence that PTEN is physically associated with platelet-derived growth factor (PDGF) receptor and that PDGF causes its dissociation from the receptor. Finally, we show that both the C2 and tail domains of PTEN contribute to binding to the PDGF receptor. These data demonstrate a novel aspect of PTEN function where it acts as an effector for the PDGF receptor function and negatively regulates PDGF receptor activation.**

- Inhibition of PTEN will promote PI3K activity and protect from reperfusion injury.

Boucher, M.; Pesant, S.; Falcao, S.; de Montigny, C.; Schampaert, E.; Cardinal, R.; Rousseau, G. Post-ischemic cardioprotection by A2A adenosine receptors: dependent of phosphatidylinositol 3-kinase pathway *J. Cardiovasc Pharmacol* 43(3) 2004 Mar, 416-22

ABSTRACT

Activation of myocardial A2A adenosine receptors during reperfusion has been shown to be cardioprotective. The intracellular mechanisms underlying this protection remain unknown. To understand the beneficial effects of activated A2A adenosine receptors in such a state, we investigated whether the enzymes phosphatidylinositol 3-kinase (PI3K) and caspase-3 can account for this post-ischemic cardioprotective effect in an anesthetized rabbit model of myocardial infarction (30 minutes ischemia; 5 hours reperfusion). Administration of the A2A agonist CGS21680 (0.2 microg/kg/min) 5 minutes before reperfusion began (Early) reduced infarct size expressed as a percentage of the area at risk (25.7 +/- 5.3% versus 46.5 +/- 5.3% for the control group; * P 60 0.05). Treatment with the A2A agonist 5 minutes after the onset of reperfusion (Late) had no effect on infarct size (38.2 +/- 6.2%). In the presence of a selective inhibitor of PI3K (>LY294002<), the beneficial effects of CGS21680 on infarct size was no longer observed (43.9 +/- 7.9%). After 5 hours of reperfusion, higher PI3K activity in the ischemic region was observed in the Early group compared with the other experimental groups. Caspase-3 activity was not observed in these

different groups. In another set of experiments, PI3K activity was significantly higher during the first 15 minutes of reperfusion in the Early group as compared with the Control group. Caspase-3 activity increased rapidly during the first 15 minutes of reperfusion in the Control group and remained stable in the Early group. These results indicated that post-ischemic cardioprotection afforded by A2A adenosine receptor activation is PI3K-dependent and modulate rapidly other signaling pathways such as caspase-3.

- PTEN and stem cells.

Our PTEN inhibitors may be able to allow stem cells to proliferate without differentiation and then use in combo with 2-(anilino)-4-aminopyrimidines types of molecules to then start differentiation

(Wu, X.; Ding, S.; Cing, Q.; Gray, N.S.; Schultz, P. G. Journal American Chemical Society 2004, 126, 1590-1591)

Stem cells are multipotent cells with the ability to self-renew and differentiate into specialized cells in response to appropriate signals.¹ Most tissues have endogenous stem/progenitor cells which, upon injury to the organ, can proliferate and differentiate at the damaged site. The adult heart is composed mainly of postmitotic and terminally differentiated cells. Although a subpopulation of myocardial cells with cardiac stem cell character was identified recently, their limited availability hinders therapeutic applications.² Stem cells derived from other tissues, such as bone marrow, have been shown to be capable of repairing heart damage in animal models,³ but inefficient differentiation and possible fusion with somatic cells limit their use in cardiac repair.⁴ Pluripotent embryonic stem (ES) cells represent a possible unlimited source of functional cardiomyocytes. However, the in vitro differentiation of ES cells into cardiomyocytes involves a poorly defined, inefficient, and relatively nonselective process.⁵ Consequently, the development of new approaches for the directed differentiation of ES cells into cardiomyocytes will likely facilitate therapeutic application of ES cells in heart disease, as well as provide important tools for probing the molecular mechanism of cardiomyocyte differentiation and heart development.

- Possible method to prevent hypoxia driven damage during and after surgery. PTEN inhibitors may enhance preconditioning in humans prior to surgery.

Carini, R; De Cesaris, MG; Splendore, R; Baldanzi, G; Nitti, MP; Alchera, E; Filigheddu, N; Domenicotti, C; Pronzato, MA; Graziani, A; Albano, E.; Role of phosphatidylinositol 3-kinase in the development of hepatocyte preconditioning. GASTROENTEROLOGY, 127 (3): 914-923 SEP 2004

Abstract:

Background & Aims: Ischemic preconditioning has been proved effective in reducing ischemia/reperfusion injury during liver surgery. However, the mechanisms involved are still poorly understood. Here, we have

investigated the role of phosphatidylinositol 3-kinase (PI3K) in the signal pathway leading to hepatic preconditioning. Methods: PI3K activation was evaluated in isolated rat hepatocytes preconditioned by 10-minute hypoxia followed by 10-minute reoxygenation. Results: Hypoxic preconditioning stimulated phosphatidylinositol-3,4,5-triphosphate production and the phosphorylation of PKB/Akt, a downstream target of PI3K. Conversely, PI3K inhibition by wortmannin or *LY294002* abolished hepatocyte tolerance against hypoxic damage induced by preconditioning. PI3K activation in preconditioned hepatocytes required the stimulation of adenosine A(2A) receptors and was mimicked by adenosine A(2A) receptors agonist CGS21680. In the cells treated with CGS21680, PI3K activation was prevented either by inhibiting adenylate cyclase and PKA with, respectively, 2,5-dideoxyadenosine and H89 or by blocking G α hi-protein and Src tyrosine kinase with, respectively, pertussis toxin and PP2. H89 also abolished the phosphorylation of adenosine A(2A) receptors. However, the direct PKA activation by forskolin failed to stimulate PI3K. This suggested that PKA-phosphorylated adenosine A(2A) receptors may activate PI3K by coupling it with G α hi-protein through Src. We also observed that, by impairing PI3K-mediated activation of phospholipase C γ (PLC γ), wortmannin and *LY294002* blocked the downstream transduction of preconditioning signals via protein kinase C (PKC) δ/ϵ isozymes. Conclusions: PI3K is activated following hepatocyte hypoxic preconditioning by the combined stimulation of adenosine A(2A) receptors, PKA, G α hi protein, and Src. By regulating PKC- ϵ/δ -dependent signals, PI3K can play a key role in the development of hepatic tolerance to hypoxia/reperfusion.

CLAIMS

1. A method of increasing neovascularization comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
2. A method of preventing cell injury comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
3. A method of preventing cell death comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
4. A method of converting cancer cells into a more treatable state comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
5. A method of converting cancer stem cells to a state whereby they can then be effectively treated with other drugs comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
6. A method of treating diabetes comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
7. A method of expanding stem cells without inducing lineage commitment comprising incubating stem cells with an effective amount of a composition comprising a PTEN inhibitor described herein.
8. A method of protecting against septic shock comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
9. A method of therapeutic angiogenesis comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
10. A method of neural stem cell self renewal comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
11. A method of preventing neurodegenerative disease comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.
12. A method of preventing apoptosis comprising administering to a patient in

need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.

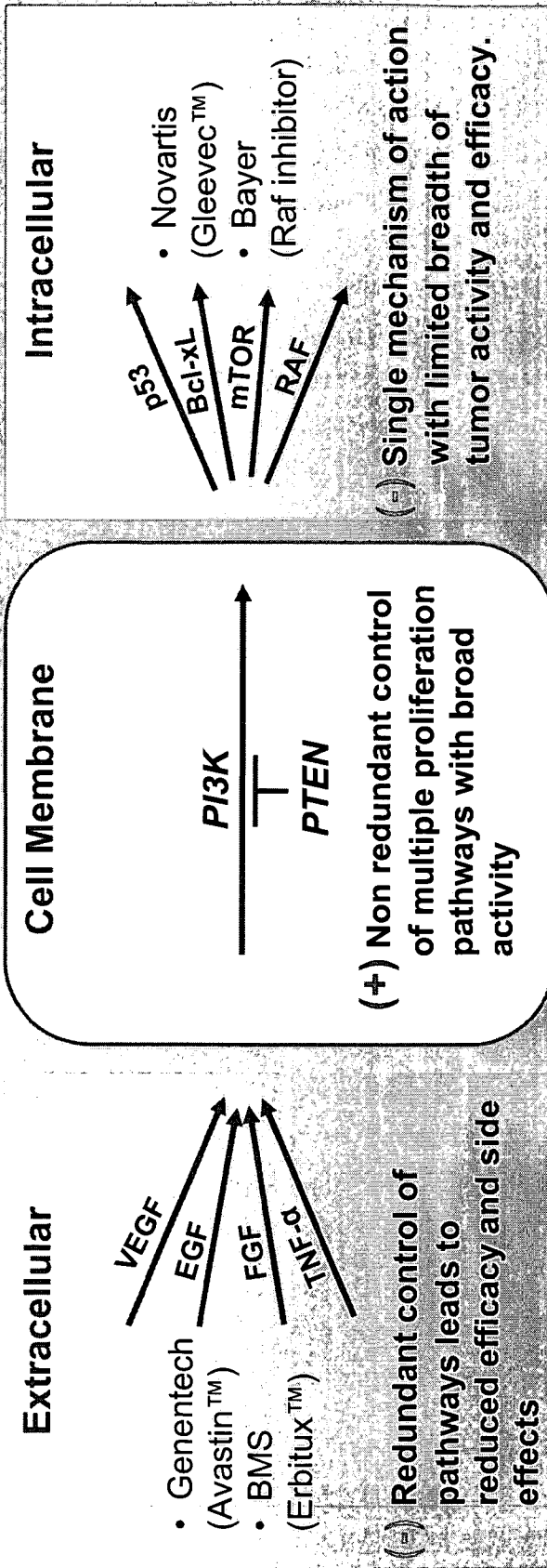
13. A method of treating obesity comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.

14. A method of blocking detrimental caspase-3 damage comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.

15. A method of preventing hypoxia driven damage comprising administering to a patient in need thereof an effective amount of a composition comprising a PTEN inhibitor described herein.

PI3K/PTEN Emerging Pathway Biology

Targeting the core **PI3K/PTEN** pathway has significant therapeutic advantages over upstream extracellular and downstream intracellular targets



■ Semafore Therapeutic Platforms

■ **PI3K Inhibition**

■ **PTEN Inhibition**

November 8, 2004

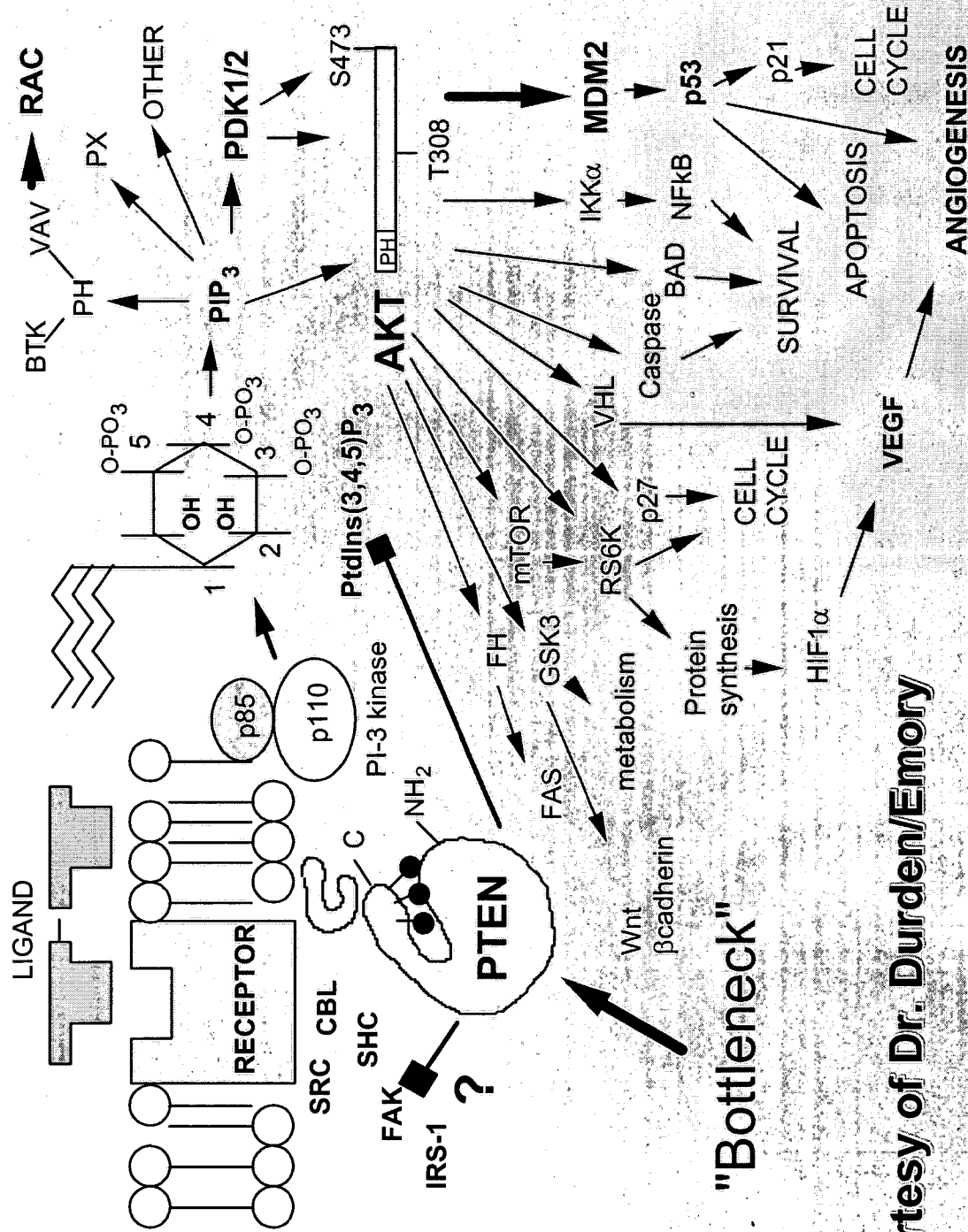
CHI: Signal Transduction

2

SEMAFORE
PHARMACEUTICALS

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PTEN/PI3K Signaling Axis Detail



Courtesy of Dr. Durden/Emory

November 8, 2004

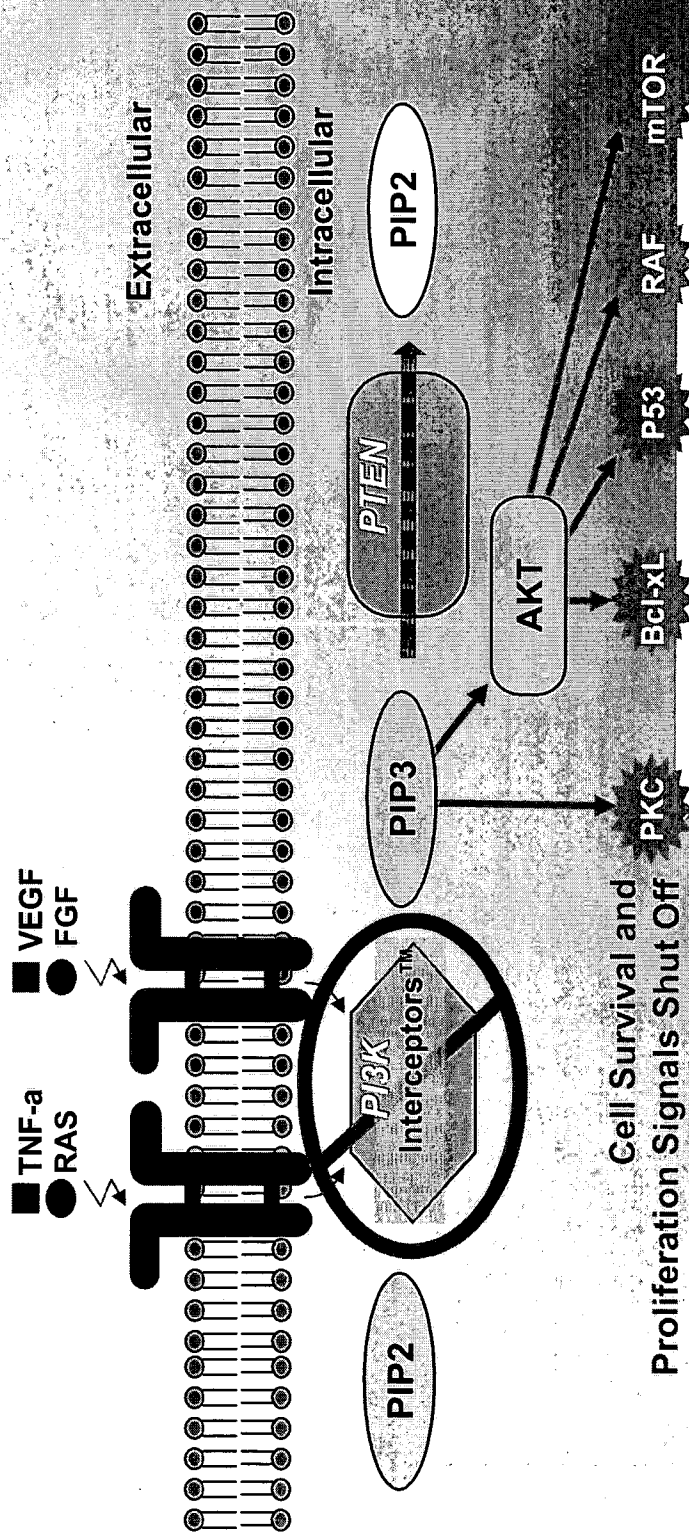
CHI: Signal Transduction

3

SEMAFORE
PHARMACEUTICALS

PI3K Inhibition Platform - Interceptors

Semafore's first core therapeutic platform is based on inhibiting the **PI3K/PTEN** pathway by inhibiting **PI3K** leading to regulated death of diseased cells



Therapeutic Applications

- Cancer
- Macular degeneration
- Inflammatory diseases
- Coronary artery disease

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Summary of PI3 Kinase Inhibitor In Vivo Studies-SF1126

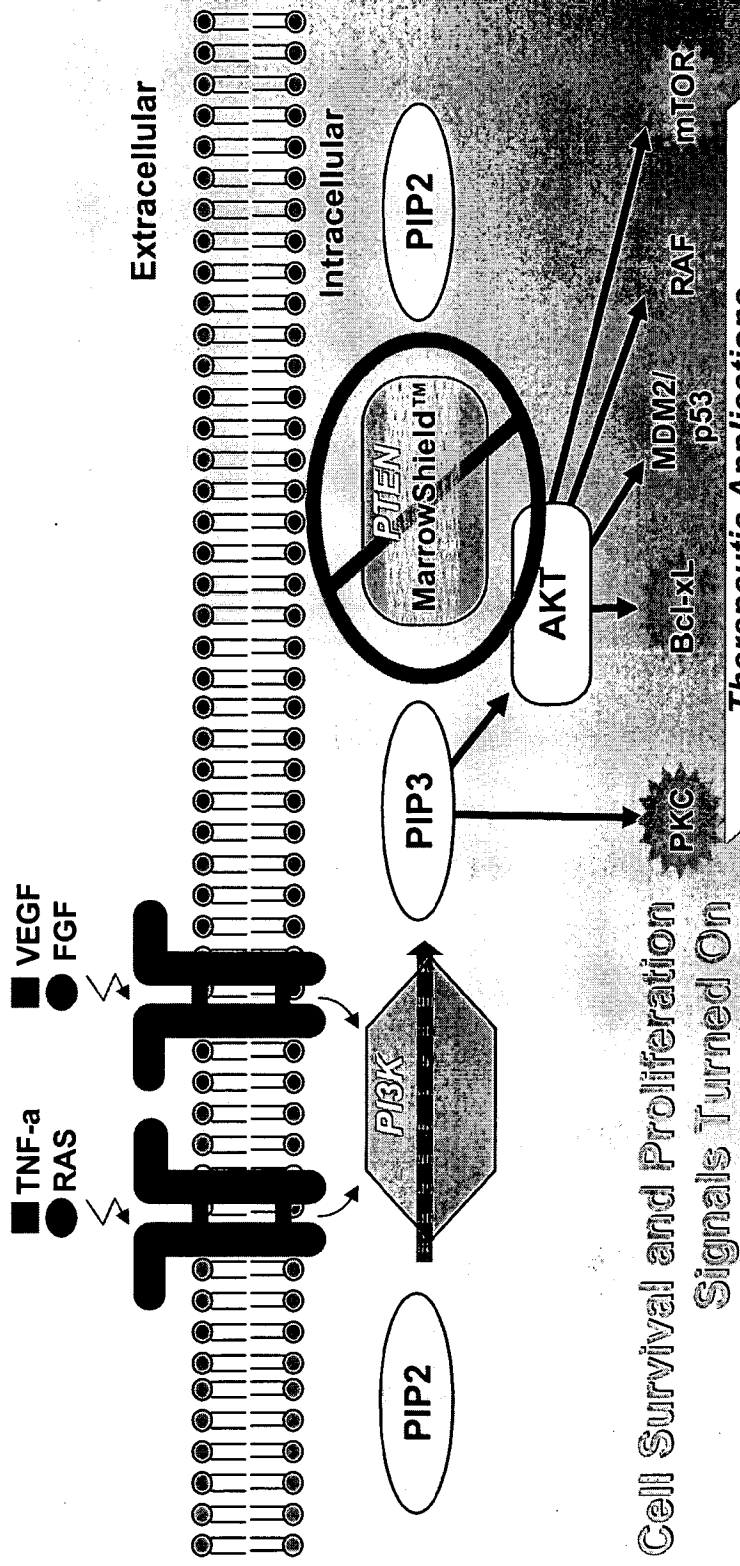
SF1126: Targeted Prodrug PI3K Inhibitor Demonstrated Xenograft Efficacy Well tolerated drug (i.v; s.c; i.m.)

| | | |
|----------|----------------------------------|-------------|
| Glioma | U87MG- PTEN null ; p53 wild type | [90% Inhib] |
| NSCL | H1299 – PTEN wild type; p53 null | [68%] |
| Prostate | PC-3 –PTEN null; p53 null | [74%] |

SF1126: Preclinical studies in progress First in man Phase I clinical-- 2005

PTEN Inhibition Platform

Semafore's second core therapeutic platform is based on promoting the **PI3K/PTEN** pathway by inhibiting **PTEN** leading to preservation of healthy disease-free cells



Therapeutic Applications

- Chemoprotection
- Radiation Protection
- Myocardial Infarction
- Stroke
- Therapeutic angiogenesis
- Stem Cell Expansion
- Diabetes
- Sepsis

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PTEN INTRODUCTION

PTEN = Phosphatase and tensin homologue deleted on chromosome 10 (MMAC)

55-kDa (403 aa) dual specificity protein (poor) and lipid phosphatase(good)
NH2-terminal catalytic domain;
COOH-terminal C2 domain
(with lipid-binding and membrane targeting functions)

Acts as a tumor suppressor by dephosphorylation of PI(3,4,5)P3 [D3 position]
(PTEN is antagonist of PI3K)

PTEN helps regulate:
cell survival
proliferation
growth
motility
angiogenesis

PTEN regulated by:

transcription
PO4 dependent stability
PDZ domain interactions
redox conditions (reversible)
(disulfide C124—C71)

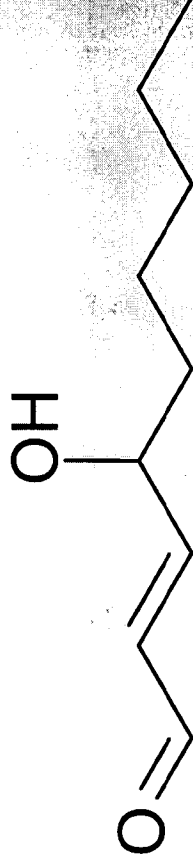
Cell Protection & Cancer Therapy

- **MarrowShield** - Single agent prophylactic for chemotherapy-induced anemia, neutropenia, and thrombocytopenia therapy; (also military applications)
- **Cancer Therapy** – novel ‘set-up & knock-down’ treatment with PI3K inhibitors [i.e. temporarily expose to PTENi to cause cancer cells to be highly addicted to PI3K pathway interruption]
- **Cardiovascular** – a) protecting needed cells from ischemia/reperfusion injury; b) stimulate angiogenesis in diseased tissue

But where are PTEN INHIBITORS?

“Specific Inhibition of PTEN Expression Reverses Hyperglycemia in Diabetic Mice”; Butler et al, Diabetes 51:1028-1034, 2002

“4-Hydroxynonenal inhibits PTEN phosphatase in vitro”; Salsman et al, Proceedings of the AACR, Vol. 44, March 2003 abstract number 3470



4-Hydroxynonenal HNE

“Bisperoxovanadium compounds are potent PTEN inhibitors” Schmid et al (Woscholski) FEBS Letters 566 (2004)35-38

PTEN Program – Semafore's Discovery Process

Semafore has developed a robust in-house **PTEN** bioassay screening program



1. In Silico –

- a. Exclusive
- b. 9 million screened
- c. Top 3000 selected
- d. Library of 100

2. In vitro – Semafore Bioassay Group

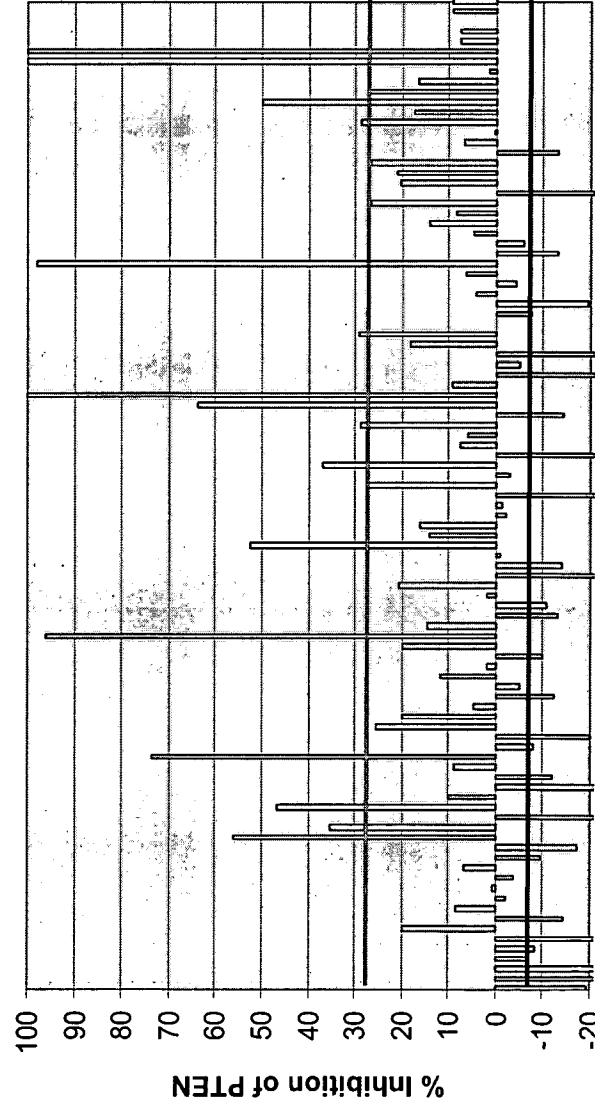
- a. 250 uM in Level 1
- b. PIP3 as substrate (PLV)

3. 40% inhibition were confirmed and rerun

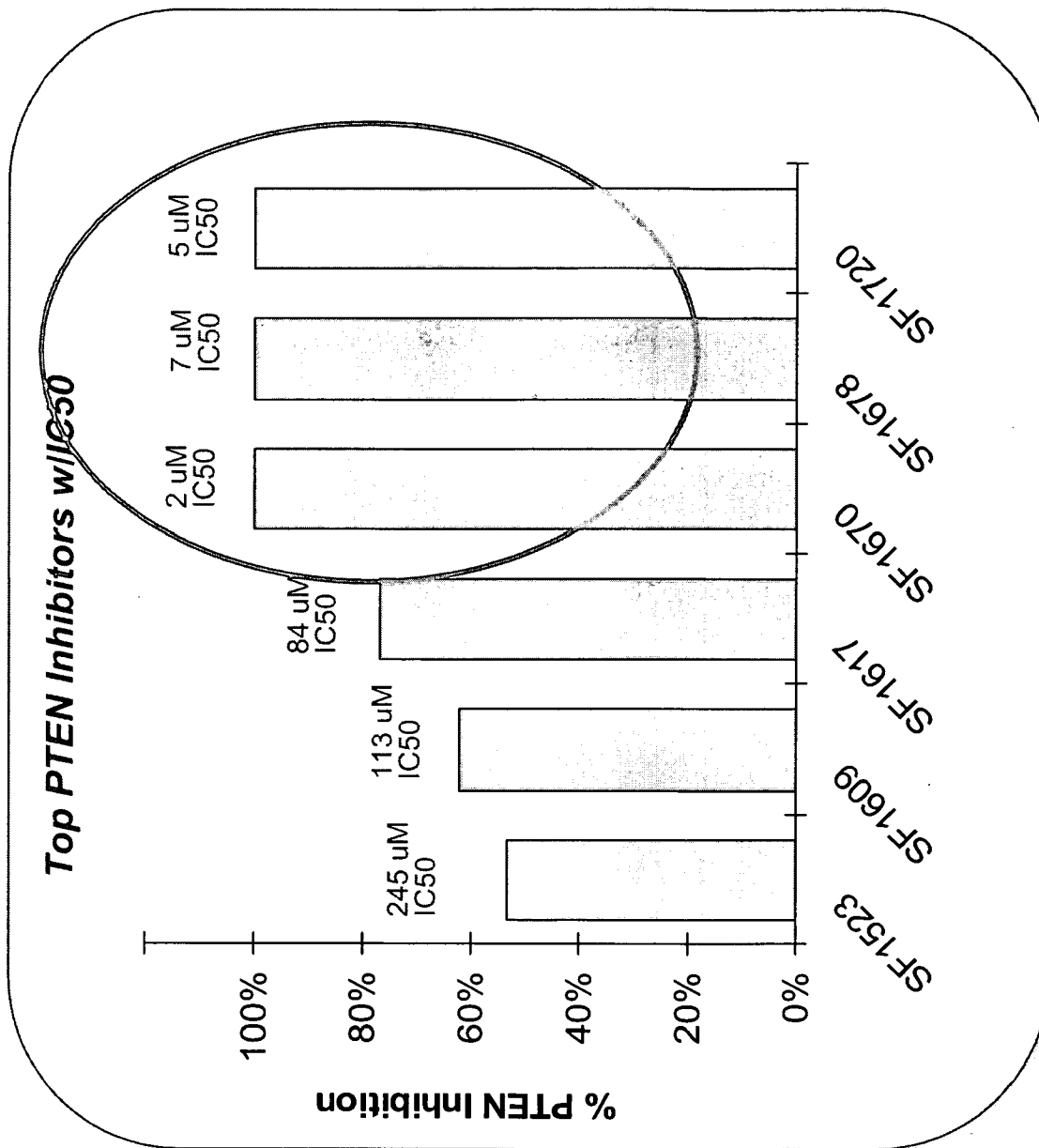
4. Enhanced Activity Noted (agonist activity)

5. IC50s determined

PTEN Assay: Initial Library Screen
%Inhibition at 250uM



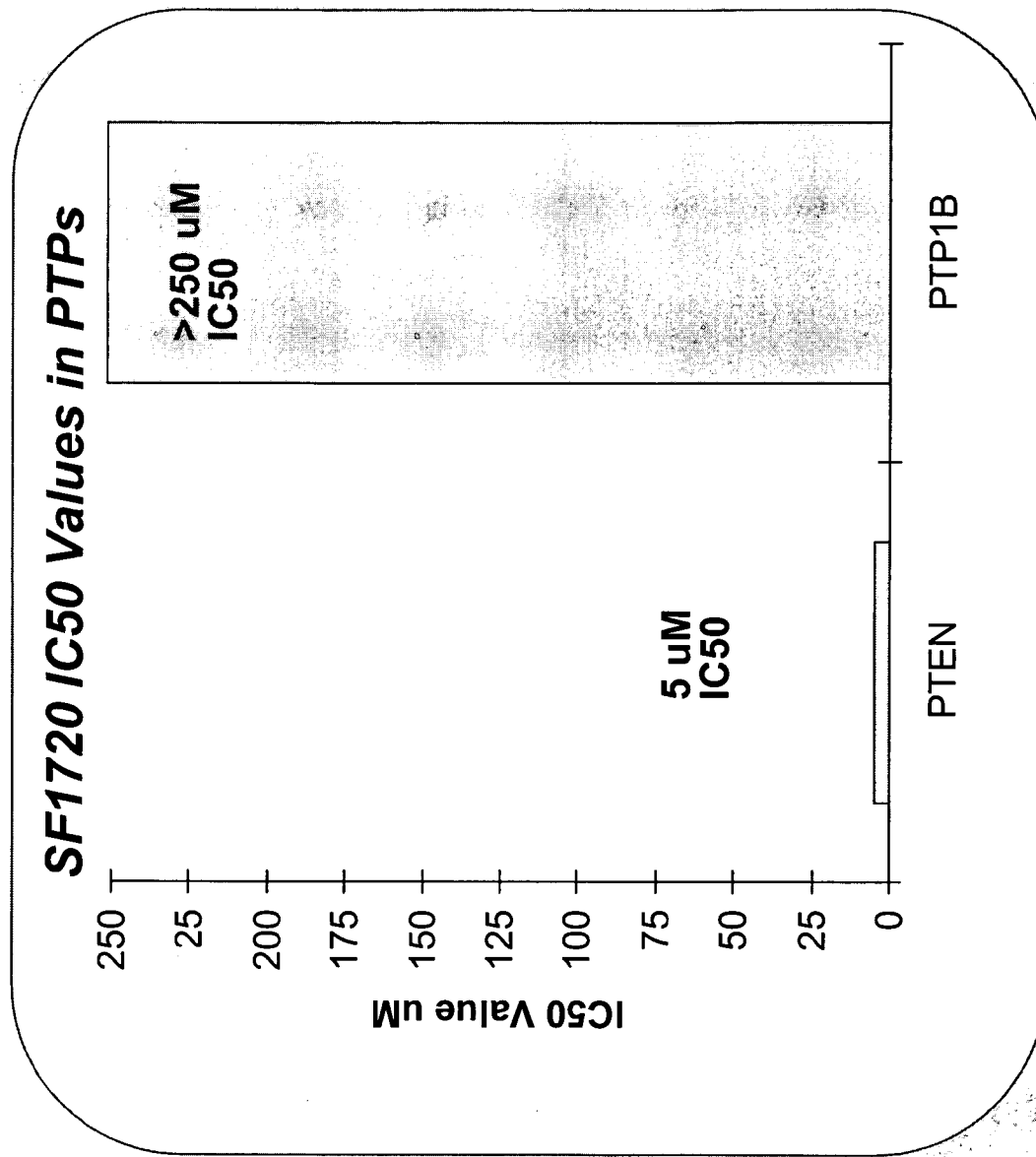
Proof Of Concept — First Known PTEN Inhibitor (Potency)



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Selectivity – SF1720 (vs PTP1B—diabetes target phosphatase)

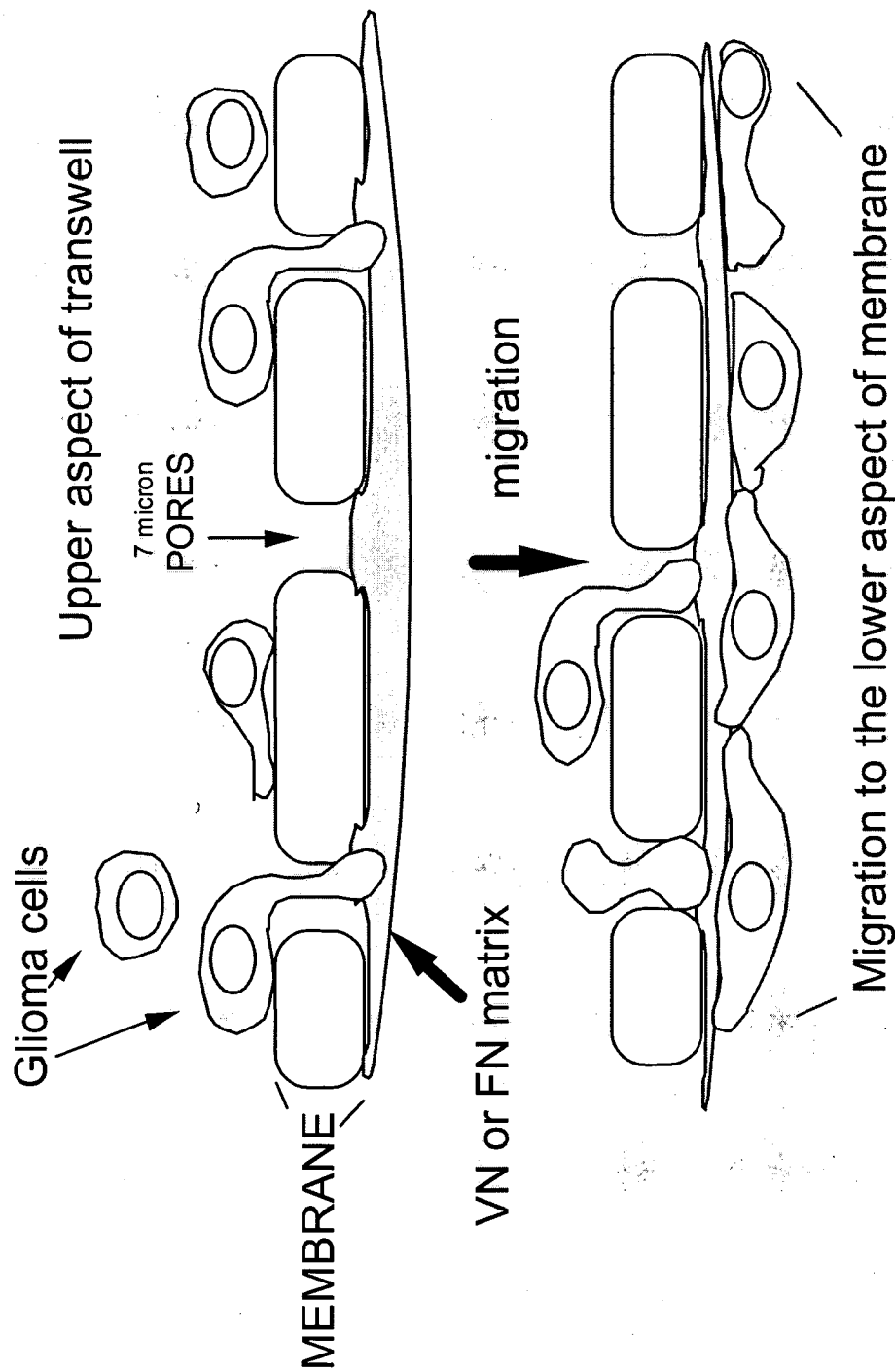


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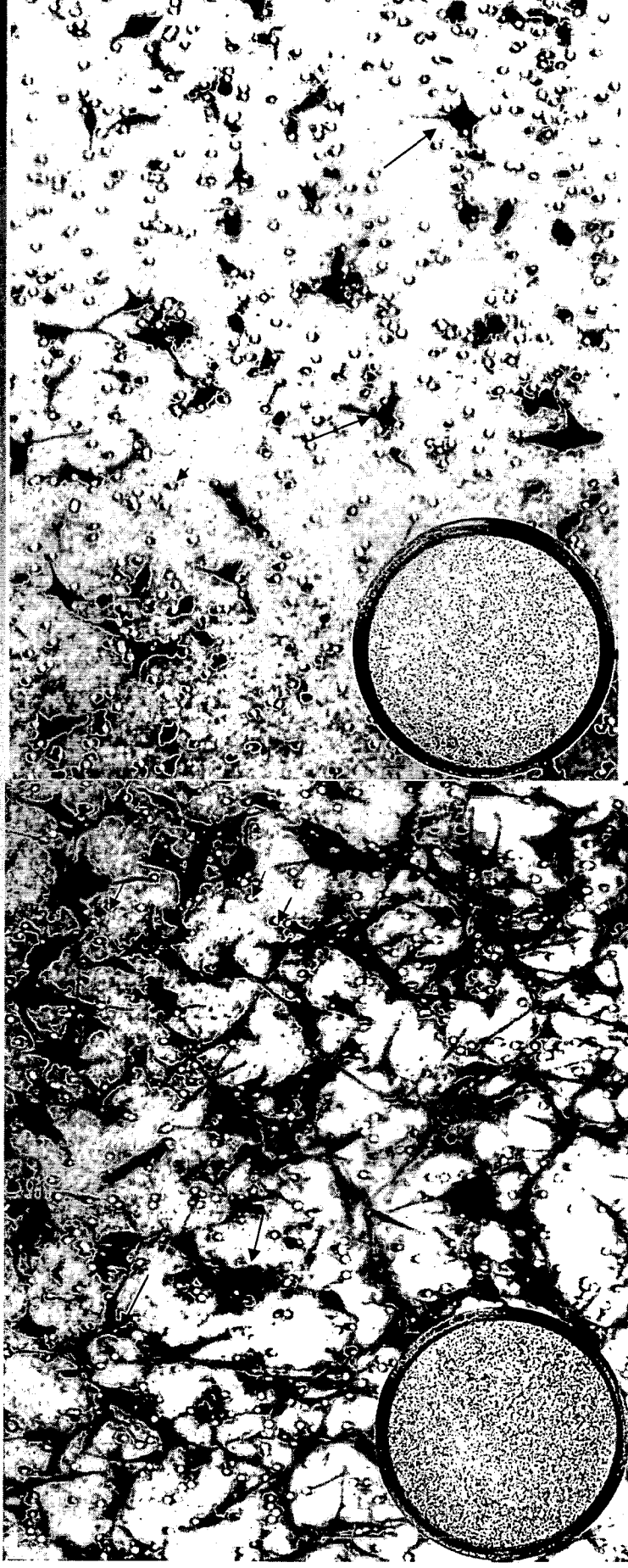
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HAPTOTAXIS ASSAY



PTEN Control of Migration

U87MG Migration on VN ($\alpha v \beta 3$)



PTEN / NULL

PTEN / WT

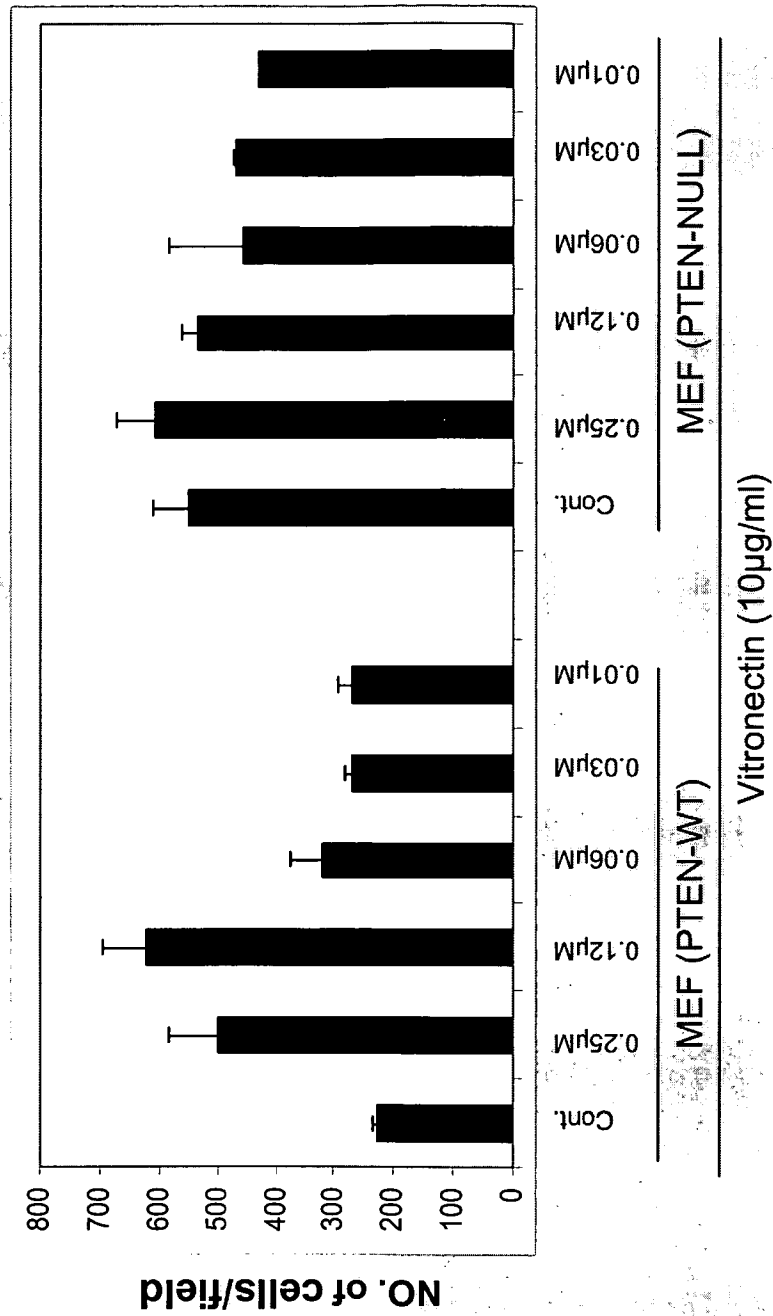
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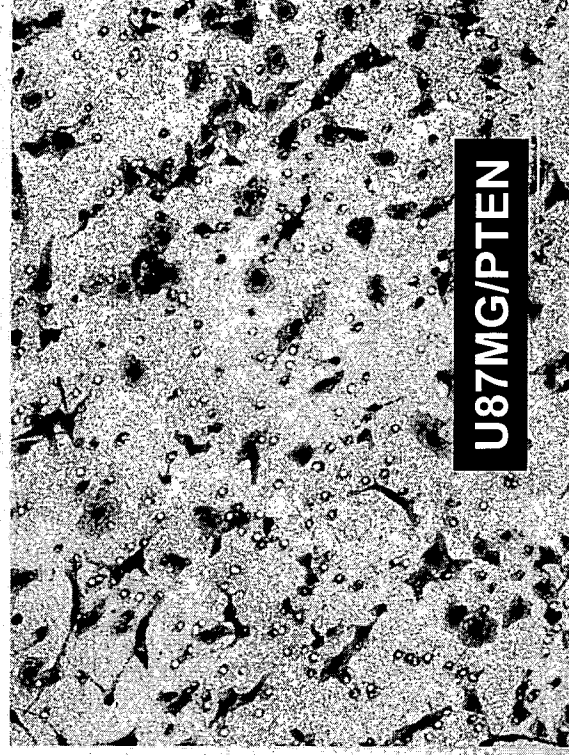
Role of PTEN inhibitor on integrin directed migration (mouse embryonic fibroblasts)



Migration Assay

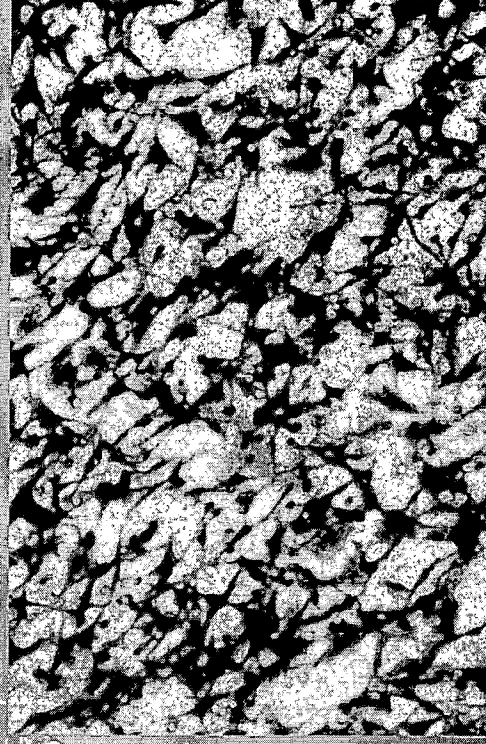


U87MG/Null



U87MG/PTEN

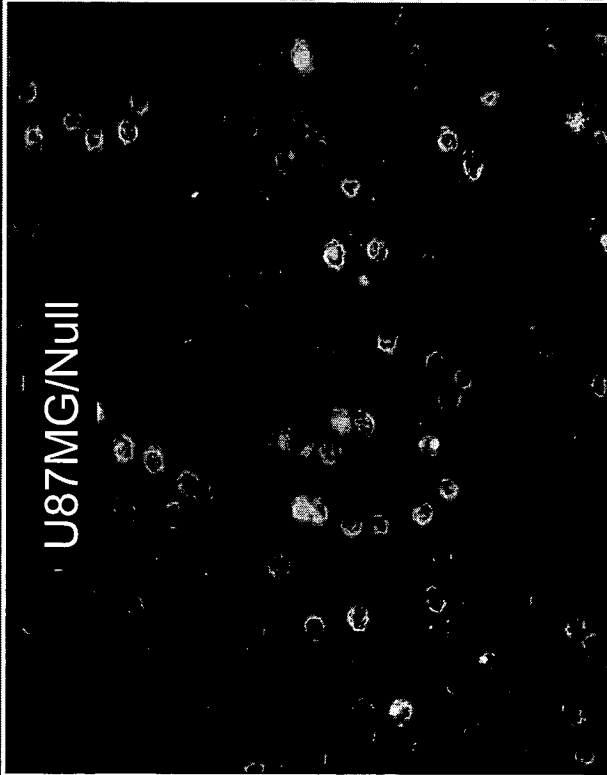
**Demonstrates Cellular Effects
of PTEN Inhibitor (SF1670)
Mimic PTEN Null Cells**



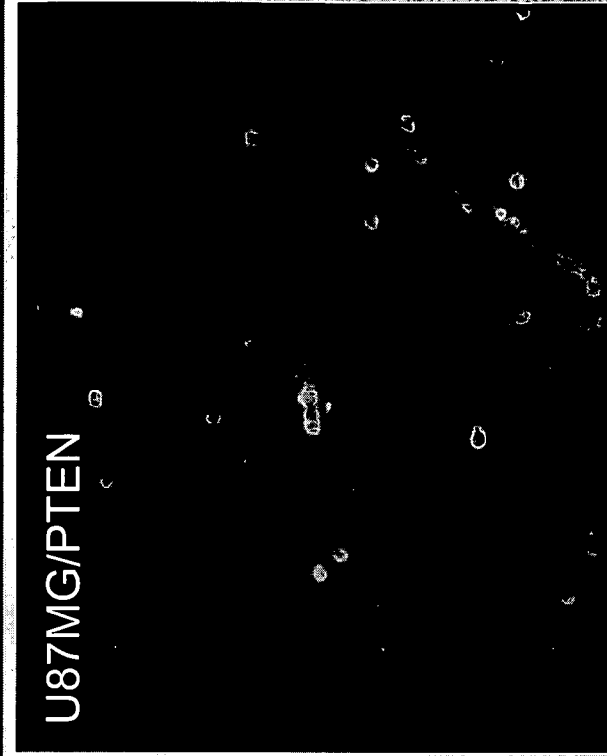
U87MG/PTEN + PTEN Inhibitor (SF1670)



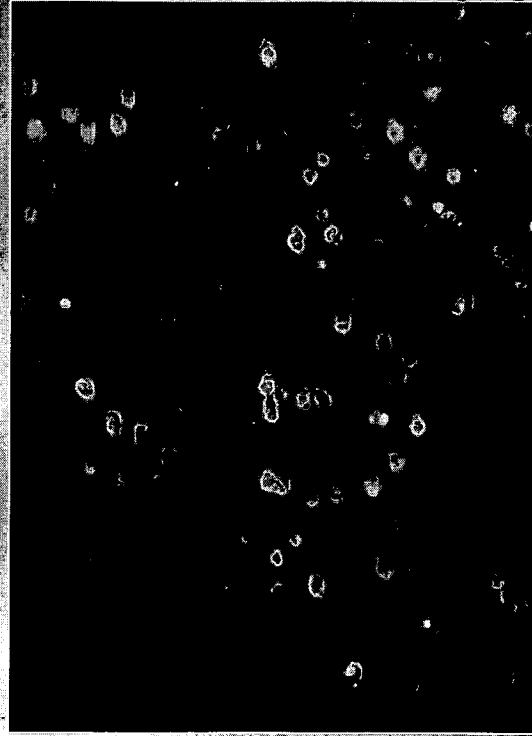
U87MG/Null



U87MG/PTEN



Merge



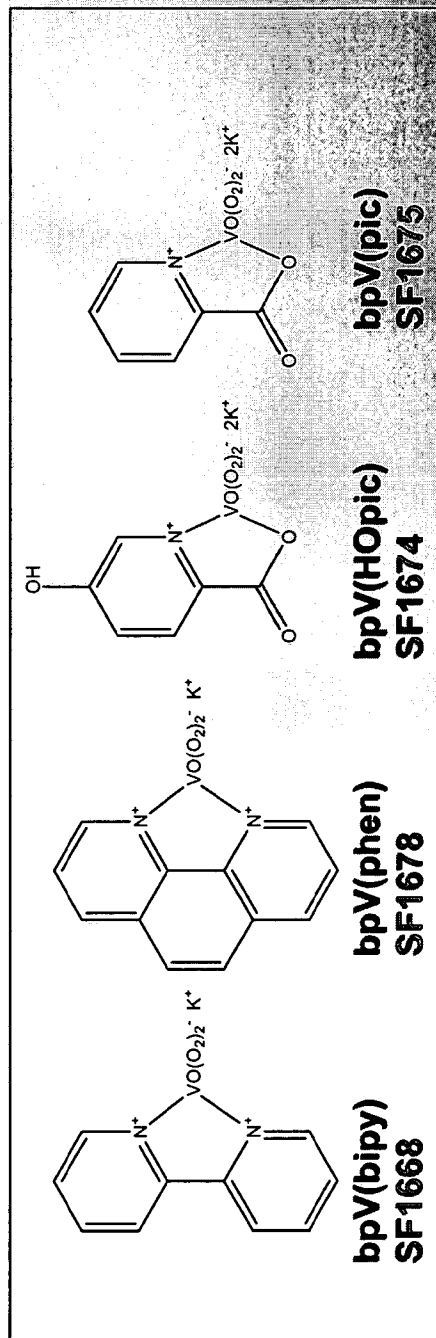
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PTEN and PTP1B Activity of Selected Vanadate Compounds

Imperial College¹ and Semafore Pharmaceuticals Assay Results

¹ Schmid, A.C; Byrne, R.D.; Vilar, R.; Woscholski, R. Bisperoxovanadium compounds are potent PTEN inhibitors, *FEBS* **2004**, 566, 35-38.

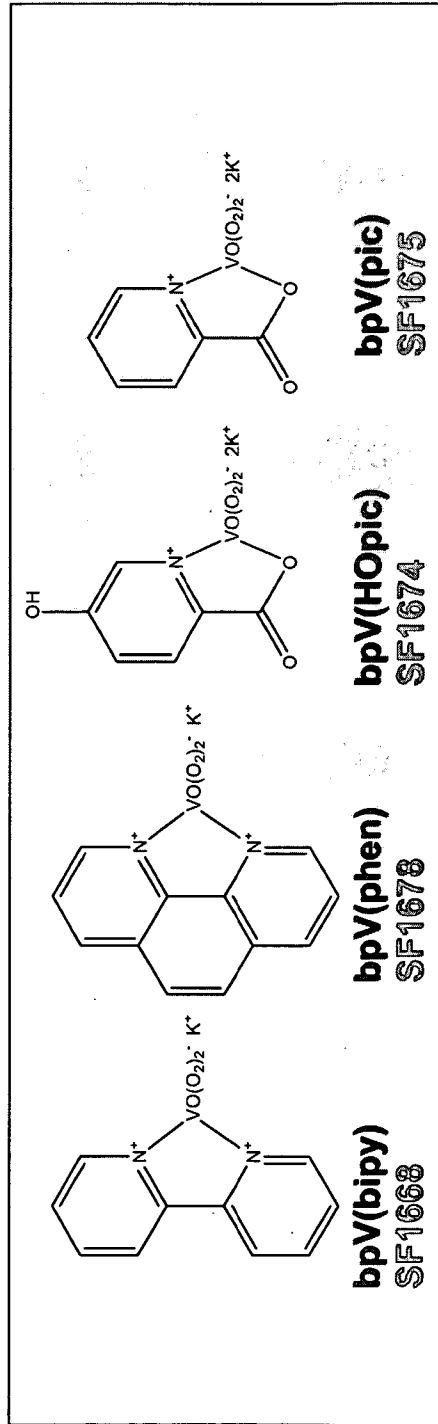


| Compound | | Imperial College ¹ | | |
|------------|--------|-------------------------------|-------------|--------------------------------|
| | | PTEN | PTP1B | Ratio (PTEN : PTP1B (pNPP)) |
| bpV(bipy) | SF1668 | 18nM ±0.8 | 164nM ±22.6 | 1:9 |
| bpV(phen) | SF1678 | 38nM ±2.4 | 920nM ±45.2 | 1:24 |
| bpV(Hopic) | SF1674 | 14nM ±2.3 | 25.3µM ±2.9 | 1:1807 |
| bpV(pic) | SF1675 | 31nM ±1.7 | | 1:1968 |
| SUBSTRATE | | PtdIns(3,4,5)P3 | pNPP | |

PTEN and PTP1B Activity of Selected Vanadate Compounds

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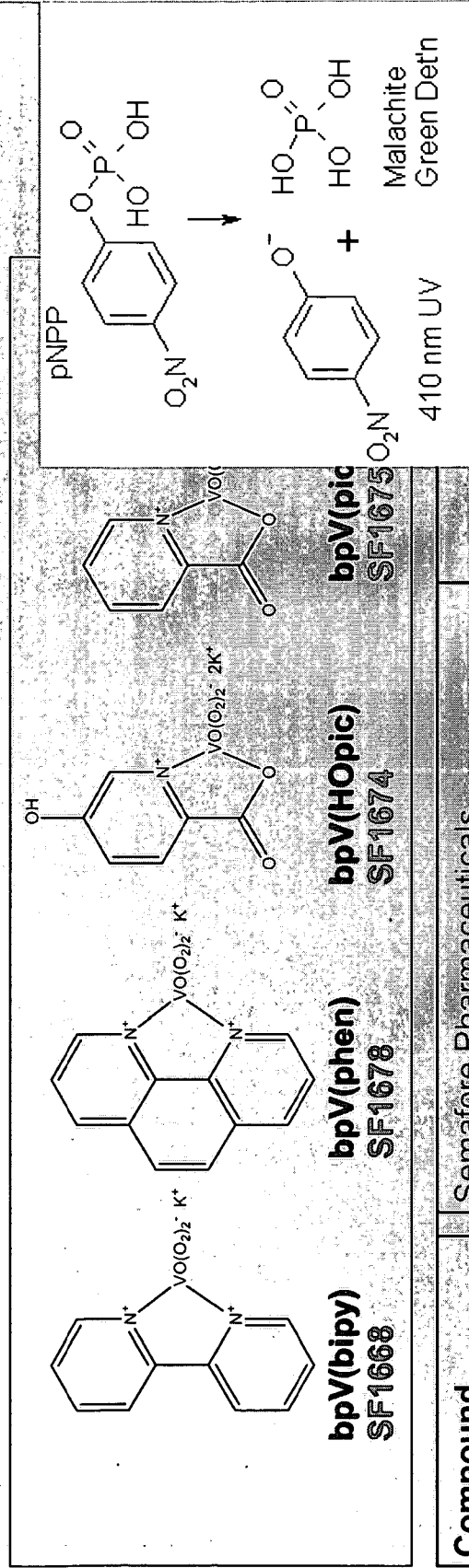


| Compound | | Semafore Pharmaceuticals | | | |
|------------|--------|--------------------------|---------------|---------|--------------------------------|
| | | PTEN | PTP1B | PTP1B | Ratio (PTEN : PTP1B (pNPP)) |
| bpV(bipy) | SF1668 | 276.3nM ±36.6 | 213.7nM ±27.1 | 103nM | 1:1 |
| bpV(phen) | SF1678 | 356.6nM ±91.4 | 97.50nM ±19.8 | 83.98nM | 0.3:1 |
| bpV(Hopic) | SF1674 | 91.1nM ± 6.4 | 79.5nM ±22.1 | 45nM | 1:1 |
| bpV(pic) | SF1675 | 111.2nM ±6.4 | 118.4nM ±6.9 | 82nM | 1:1 |
| SUBSTRATE | | PtdIns(3,4,5)P3 | pNPP | GluTyr | |

PTEN and PTP1B Activity of Selected Vanadate Compounds

Imperial College¹ and Semafore Pharmaceuticals Assay Results

¹ Schmid, A.C.; Byrne, R.D.; Vilar, R.; Woscholski, R. Bisperoxovanadium compounds are potent PTEN inhibitors, *FEBS* **2004**, 566, 35-38.

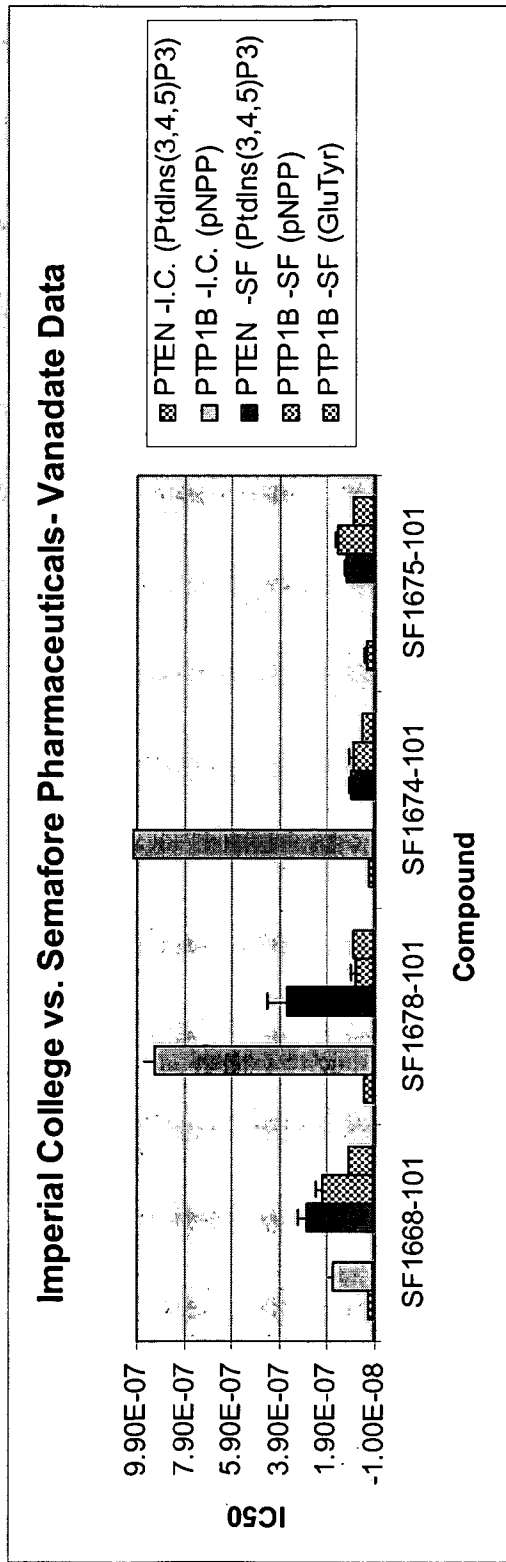


| Compound | Semafore Pharmaceuticals | | | | Ratio (PTEN : PTP1B (pNPP)) |
|-------------------|--------------------------|---------------|---------|--|--------------------------------|
| | PTEN | PTP1B | PTP1B | | |
| bpV(bipy) SF1668 | 276.3nM ±36.6 | 213.7nM ±27.1 | 103nM | | 1:1 |
| bpV(phen) SF1678 | 356.6nM ±91.4 | 97.50nM ±19.8 | 83.98nM | | 0.3:1 |
| bpV(Hopic) SF1674 | 91.1nM ±6.4 | 79.5nM ±22.1 | 45nM | | 1:1 |
| bpV(pic) SF1675 | 111.2nM ±6.4 | 118.4nM ±6.9 | 82nM | | 1:1 |
| SUBSTRATE | PtdIns(3,4,5)P3 | pNPP | GluTyr | | |

PTEN and PTP1B Activity of Selected Vanadate Compounds

Imperial College¹ and Semafore Pharmaceuticals Assay Results

- PTEN inhibitors have triple digit nanomolar inhibition, comparable to our SF compounds
- PTP1B inhibitors have triple digit nanomolar inhibition (no selectivity)
- PTP1B substrate- GluTyr vs pNpp (dynamic range, 410nM vs. 620 nM)



| Compound | Imperial College ¹ | Semafore Pharmaceuticals |
|------------|-------------------------------|-----------------------------|
| | | Ratio (PTEN : PTP1B (pNPP)) |
| bpV(bipy) | SF1668 | 1:9 |
| bpV(phen) | SF1678 | 1:24 |
| bpV(Hopic) | SF1674 | 1:1807 |
| bpV(pic) | SF1675 | 1:1968 |

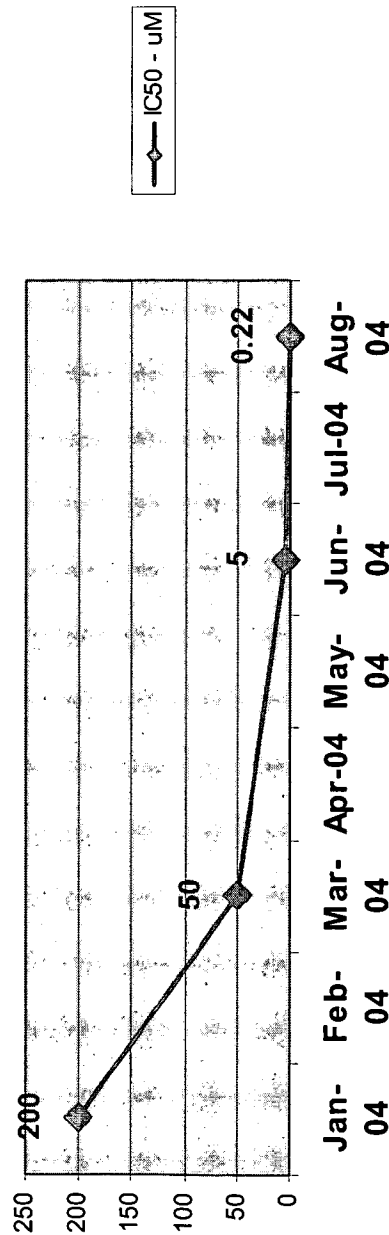
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PTEN Inhibitor Optimization – NanoMolar, Selective

Semafore has optimized PTEN inhibitors from >200uM to less than 300 nM, with increasing selectivity vs. other phosphatases.

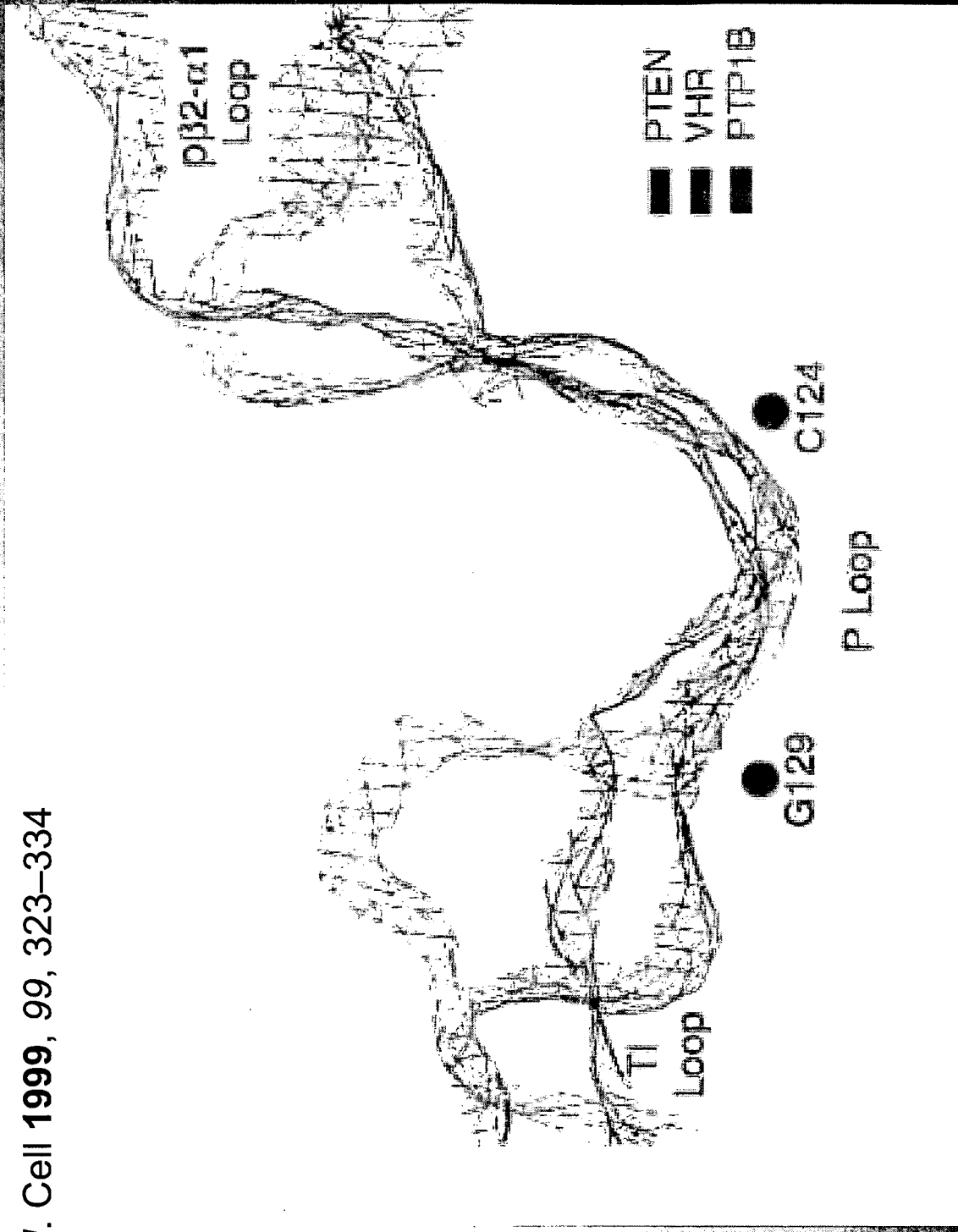
IC50 Progression of PTEN Inhibitor Program



SF1751 = 200nM, 10x Selectivity

Selective PTEN Inhibitors-- Focus on Pocket Size and Shape

Lee et al. Cell 1999, 99, 323-334



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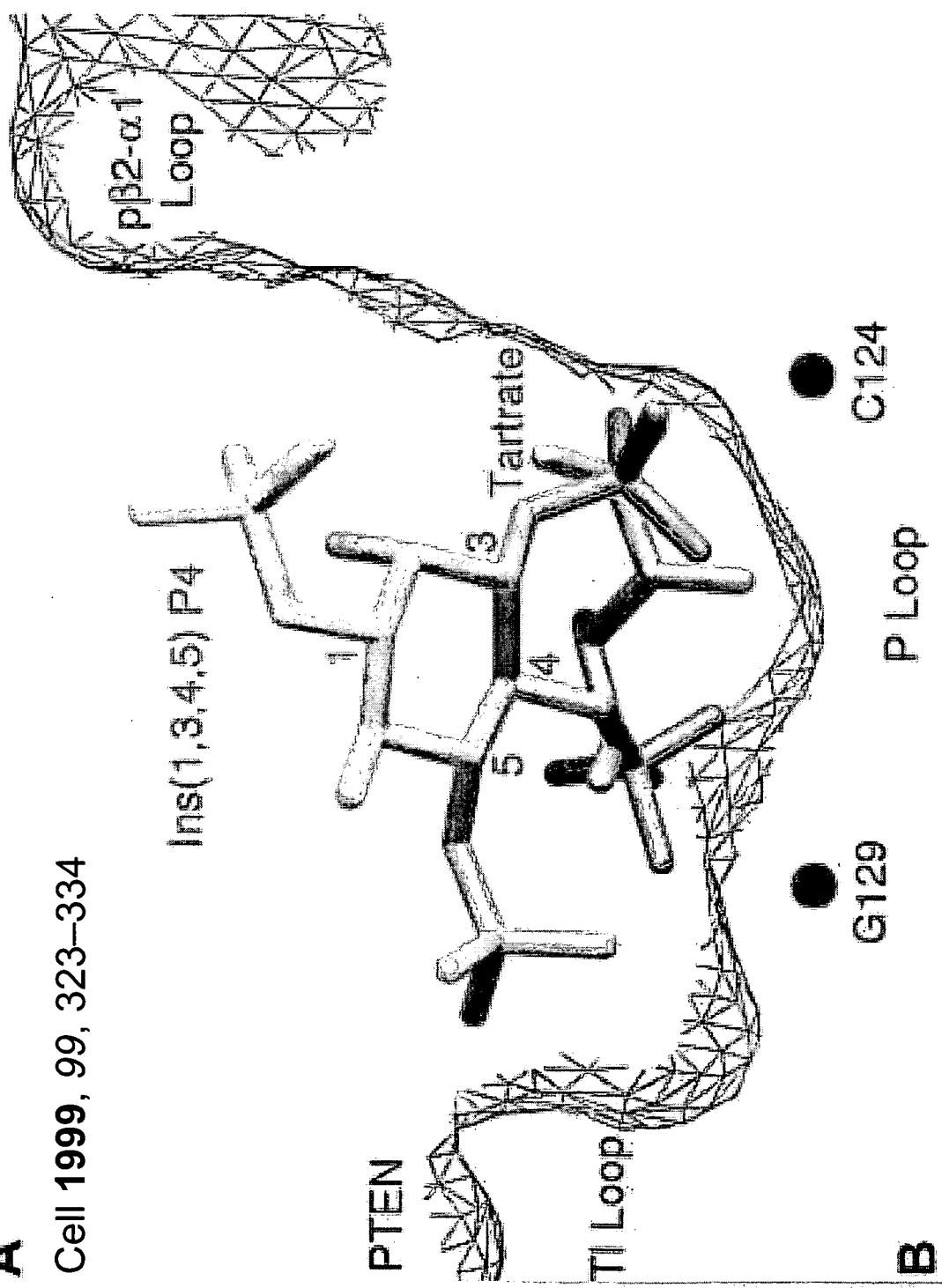
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Selective PTEN Inhibitors-- Focus on Pocket Size and Shape

Lee et al. Cell 1999, 99, 323-334

A



B

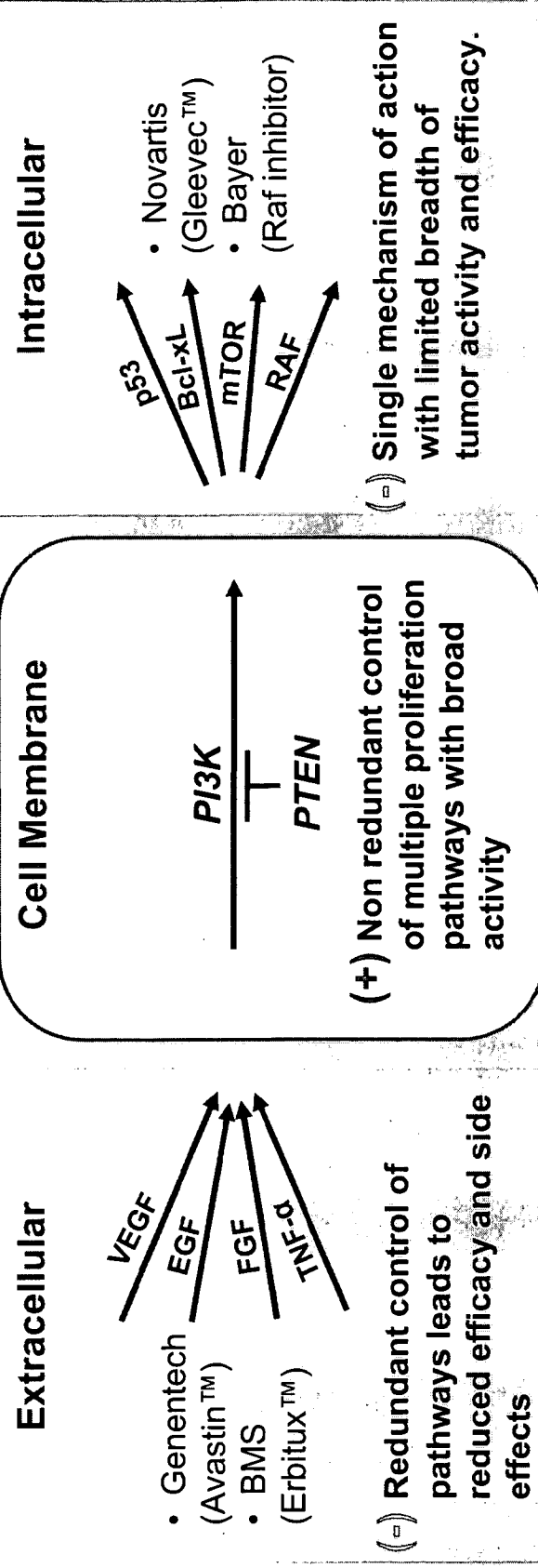
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SUMMARY



■ Semafore Therapeutic Platforms

■ *PI3K Inhibition—SF1126 PRECLINICAL*

■ *PTEN Inhibition—Potency good; refining selectivity*

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